**EXHIBIT B** 

POLYPEPTIDE HAVING AN ACTIVITY TO SUPPORT PROLIFERATION
OR SURVIVAL OF HEMATOPOIETIC STEM CELL AND HEMATOPOIETIC
PROGENITOR CELL, AND DNA CODING FOR THE SAME

## Background of the Invention

## Field of the Invention

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The present invention relates to a polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, a DNA coding the polypeptide, and a pharmaceutical composition comprising the polypeptide as active ingredient.

## Description of the Related Art

15 Fully differentiated mature hematopoietic cells have limited short lives. Homeostasis of the blood is maintained due to supply of the mature blood cells caused by continuous differentiation of hematopoietic progenitor cells. The hematopoietic progenitor cells 20 are giving rise from more undifferentiated hematopoietic stem cells. The hematopoietic stem cells have potential of differentiating into all of the differentiation lineages (totipotency) and have potential of self-renew with retaining the totipotency 25 so as to supply the hematopoietic cells through life. That is, the hematopoietic stem cells are known to generate totipotent stem cells by the self-renew and to differentiate in parts to a variety of the mature blood cells through the hematopoietic progenitor cells.

This differentiation of the blood cells is regulated by a variety of cytokines. Erythropoietin is known to 5 promote the differentiation of the erythrocytic lineages. G-CSF and thrombopoietin are also known to promote the differentiation of the neutrophils, and the megakaryocytes and the platelet productive cells, respectively. However, a factor required for the self-10 renew of the hematopoietic stem cell with retaining the totipotency has not been clear. Although SCF/MGF (Williams, D.E., Cell, 63: 167-174, 1990; Zsebo, K.M., Cell, 63: 213-224, 1990), SCGF (WO98/08869), and the like are reported as growth factors for the hematopoietic stem cells, none of them have potency to 15 sufficiently retain the totipotency of the hematopoietic stem cells. Although attempts to culture the hematopoietic stem cells in the presence of combinations of known cytokines, a system for efficient amplification of the hematopoietic stem cells was not realized (Miller, 20 C. L., Proc. Natl. Acad. Sci. USA, 94: 13648-13653, 1997; Yaqi, M., Proc. Natl. Acad. Sci. USA, 96: 8126-8131, 1999; Shih, C.C., Blood, 94: 5 1623-1636, 1999).

On the other hand, attempts to allow the

hematopoietic stem cells to survive or proliferate

without differentiation by using stromal cells which

supply an environment suitable for survival or

proliferation of the hematopoietic stem cells were reported (Moore K.A., Blood, 89: 12, 4337-4347, 1997). In addition, WO99/03980 discloses a stromal cell line capable of supporting proliferation or survival of hematopoietic stem cells and hematopoietic progenitor cells, which are established from an AGM (Aorta-Gonad-Mesonephros) region of a fetal mouse.

It is postulated that there should be more peptides that efficiently facilitate hematopoietic stem cell and progenitor cell amplification by themselves or in combination with stromal cells or stimulating factors such as cytokines, in addition to known factors affecting hematopoietic cells.

# 15 Summary of the Invention

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Since the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells in vitro can be supported by co-culture of stromal cells and hematopoietic stem cells and hematopoietic progenitor cells, the stromal cells are expected to produce factors supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells. An object of the present invention is to provide a factor supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, which is derived from the stromal cells.

The inventor of the present invention has assumed

that the mouse stromal cells produce factors supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, as mentioned above. Attention is given that there are two kinds of stromal cells. One has a ability to support the 5 proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells (hereafter sometimes referred to as "activity to support hematopoietic stem cells"). The other does not have the activity to 10 support hematopoietic stem cells. The inventor of the present invention has assumed that this difference in the ability is due to the fact that expression of genes encoding the factors is increased in the supporting stromal cells and that the expression is low in non-15 supporting stromal cells. Thus the inventor think it can be found the factors supporting the proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells among the genes expressed higher in the supporting cells compared to in the non-supporting cells. 20 In this context, the inventor has identified genes of which expressions are high in AGM-s3-A9 cell line which has the activity to support hematopoietic stem cells, and low or undetected in AGM-s3-A7 cell line which does not have the activity to support hematopoietic stem 25 cells, and has determined the activities to support hematopoietic stem cells, of cells in which these gene groups are highly expressed. As a result, the present

invention has been completed.

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That is, the present invention provides the followings.

- (1) A DNA coding for a polypeptide of the
  5 following (A) or (B):
  - (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25; or
- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- 15 (2) The DNA according to (1), which is a DNA of the following (a) or (b):
  - (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the nucleotide sequence of nucleotides 642 to 1370 of SEQ ID NO: 22, and the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24; or
  - (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic

progenitor cells.

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- (3) The DNA according to (2), the stringent condition is 6 x SSC, 5 x Denhardt, 0.5% SDS and 68°C (SSC: 3 M NaCl, 0.3 M sodium citrate; 50 x Denhardt: 1% BSA, 1% polyvinyl pyrrolidone, 1% Ficoll 400), or 6 x SSC, 5 x Denhardt, 0.5% SDS, 50% formamide and 42°C.
- (4) A expression vector which comprises the DNA of any one of (1) to (3) in such a manner that the DNA can be expressed.
- 10 (5) A cell into which the DNA of any one of (1) to (3) is introduced in such a manner that the DNA can be expressed.
  - (6) A polypeptide which is an expression product of the DNA of any one of (1) to (3), the polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
  - (7) The polypeptide according to (6), which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25, or an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence.
- (8) The polypeptide according to (6) or (7),
  25 which is modified with one or more modifying agents selected from the group consisting of polyethylene glycol (PEG), dextran, poly(N-vinyl-pyrrolidone),

polypropylene glycol homopoymer, copolymer of polypropylene oxide/ethylene oxide, polyoxyethylated polyol and polyvinyl alcohol.

(9) An monoclonal antibody which binds to the polypeptide of any one of (6) to (8).

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- (10) A method for supporting proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, comprising the step of co-culturing stromal cells in which a DNA coding for a polypeptide of the following (A) or (B) is expressed, with hematopoietic stem cells or progenitor cells,
- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- (11) The method according to (10), wherein the DNA is a DNA of the following (a) or (b):
- 25 (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide sequence of nucleotides 1 to 1671 of SEQ ID NO: 8, the

nucleotide sequence of nucleotides 1 to 1674 of SEQ ID NO: 10, the nucleotide sequence of nucleotides 1 to 366 of SEQ ID NO: 12, the nucleotide sequence of nucleotides 84 to 1121 of SEQ ID NO: 14, the nucleotide sequence of nucleotides 1 to 1035 of SEQ ID NO: 16, the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 20, the nucleotide sequence of nucleotides 642 to 1370 of SEQ ID NO: 22, the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24, the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24, the nucleotide sequence of nucleotide sequence of nucleotides 1 to 2487 of SEQ ID NO: 26, and the nucleotide sequence of nucleotides 1 to 2496 of SEQ ID NO: 28; or

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- (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- 20 (12) A method for supporting proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, comprising the step of culturing hematopoietic stem cells or progenitor cells in the presence of a polypeptide of the following (A) or (B), said polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells when the hematopoietic

stem cells or hematopoietic progenitor cells are cultured in the presence of the polypeptide,

- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- effect to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, which comprises an effective amount of a polypeptide of the following (A) or (B), said polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells when hematopoietic stem cells or hematopoietic progenitor cells are cultured in the presence of the polypeptide,
- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 21, SEQ ID NO: 23,

SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or

(B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.

Terms used in this specification are defined as follows.

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A hematopoietic stem cell is defined as a cell 10 having totipotency, that is, ability to differentiate into all the cell lineages of the blood cells, and having a potency of self-renew with retaining the totipotency. A hematopoietic progenitor cell is defined as a cell which can differentiate a single cell lineage 15 of the blood cell or plural cell lineages but cannot differentiate into all of the cell lineages. A stromal cell is defined as a cell which can be co-cultured together with the hematopoietic stem cells to construct 20 a hematopoietic environment simulating in vivo hematopoietic environment in vitro. Cells derived from any origin can be used as long as the cells can be cocultured with the hematopoietic cells in vitro.

Erythrocyte progenitor cells hardly survive and proliferate in *in vitro* culture environments and rapidly disappear. If the survival and proliferation of the erythrocyte progenitor cells are observed, continuous

production of the erythrocyte progenitor cells is

predicted to occur due to the survival and proliferation

of the more immature hematopoietic stem cells or the

hematopoietic progenitor cells. Therefore, in an

5 assessment system of human hematopoietic stem cells,

proliferation of hematopoietic stem cells or immature

hematopoietic progenitor cells can be determined by

using the survival and proliferation of the erythrocyte

progenitor cells (BFU-E, CFU-E, and CFU-E mix) as an

10 index.

# Brief Explanation of the Drawings

Fig. 1 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-s3 subclone A9, A7, or D11 cells for two weeks.

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Fig. 2 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-s3 subclone A9, A7, or OP9 cells for two weeks.

Fig. 3 shows time course of donor derived lymphoid lineage cells or myeloid lineage cells reconstitution in irradiated recipient mice that received the hematopoietic stem cells co-cultured with stromal cells.

Fig. 4 shows proliferation statuses of hematopoietic

stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-2 is highly expressed (A9/SCR-2), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

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Fig. 5 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A7 cells in which a gene SCR-2 is highly expressed (A7/SCR-2), AGM-S3-A7 cells into which a control vector is introduced (A7/pMXIG) or AGM-S3-A7 cells (A7) for two weeks.

Fig. 6 shows time course of donor derived lymphoid

lineage cells or myeloid lineage cells reconstitution in

peripheral blood of irradiated recipient mice that

received the hematopoietic stem cells co-cultured with

AGM-S3-A7 cells in which a gene SCR-3 is highly

expressed (A7/SCR-3), AGM-S3-A7 cells into which a

control vector is introduced (A7/pMXIG) or AGM-S3-A7

cells.

Fig. 7 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-4 is highly expressed (A9/SCR-4), AGM-S3-A9 cells into which a control vector is introduced

(A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

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Fig. 8 shows time course of donor derived lymphoid lineage cells or myeloid lineage cells reconstitution in peripheral blood of irradiated recipient mice that received the hematopoietic stem cells co-cultured with AGM-S3-A7 cells in which a gene SCR-5 is highly expressed (A7/SCR-5), AGM-S3-A7 cells into which a control vector is introduced (A7/pMXIG) or AGM-S3-A7 cells.

Fig. 9 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-6 is highly expressed (A9/SCR-6), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

Fig. 10 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-7 is highly expressed (A9/SCR-7), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

25 Fig. 11 shows proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells determined by a clonogenic assay after co-culture

of CD34-positive hematopoietic stem cells with AGM-S3-A9 cells in which a gene SCR-8 is highly expressed (A9/SCR-8), AGM-S3-A9 cells into which a control vector is introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks.

# Detailed Description of the Invention

Hereafter, the present invention will be described in detail below.

The following genes are those identified as genes of which expressions are high in AGM-s3-A9 cell line which has the activity to support hematopoietic stem cells, and low or undetected in AGM-s3-A7 cell line which does not have the activity to support hematopoietic stem cells, and determined to have the activities to support hematopoietic stem cells, of cells in which these gene groups are highly expressed.

Gene SCR-2

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The gene is the same gene as a mouse gene, Mus musculus glypican-1 (GPC-1) of a GenBank accession number AF185613.

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 8. Only the amino acid sequence is shown in SEQ ID NO: 9.

The human amino acid sequence of GPC-1 is recorded

in GenBank under an accession number P35052, and the human nucleotide sequence of GPC-1 is recorded in GenBank database under an accession number AX020122. It is predicted that the similar activity is detected in the human gene.

The nucleotide sequence of the gene from human and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 10. Only the amino acid sequence is shown in SEQ ID NO: 11.

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existing on a cell surface, and have a characteristic structure such as cysteine rich globular domain, short glycosaminoglycan binding domain, glycosylphosphatidylinositol membrane binding domain. Six family genes from glypican-1 to glypican-6 have been found (J Biol Chem 1999 Sep 17;274(38):26968-77, Glypican-6, a new member of the glypican family of cell surface heparan sulfate proteoglycans. Veugelers M, De Cat B, Ceulemans H, Bruystens AM, Coomans C, Durr J, Vermeesch J, Marynen P, David G).

With respect to biological activities of GPC-1, there are a number of reports: To regulate growth stimulating activity of heparin binding growth factors (fibroblast growth factor 2 (FGF2), heparin-binding EGF-like growth factor (HB-EGF)) to promote proliferation of cancer cells showing autocrine proliferation by stimulation by the growth factors (J Clin Invest 1998)

Nov 1; 102(9):1662-73, The cell-surface heparan sulfate proteoglycan glypican-1 regulates growth factor action in pancreatic carcinoma cells and is overexpressed in human pancreatic cancer., Kleeff J, Ishiwata T, Kumbasar A, Friess H, Buchler MW, Lander AD, Korc M).

To bind HGF (hepatocyte groth factor) to promote reactivity with cytokines, of antigen-specific B cells. To participate in association of a cell with an adhesive molecule to involve in invasion of the cell (J Biol Chem 10 1998 Aug 28;273(35):22825-32, Heparan sulfate proteoglycans as adhesive and anti-invasive molecules. Syndecans and glypican have distinct functions., Liu W, Litwack ED, Stanley MJ, Langford JK, Lander AD, Sanderson RD). These findings show that GPC-1 involves 15 in activity expression of various cell-stimulating factors. Also, there is a report that expression of the glypican family gene in bone marrow is confirmed (Biochem J 1999 Nov 1;343 Pt 3:663-8, Expression of proteoglycan core proteins in human bone marrow stroma., 20 Schofield KP, Gallagher JT, David G). However, in these reports, it is not described about effects of GPC-1 on hematopoietic stem cells or hematopoietic progenitor cells.

## 25 Gene SCR-3

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The gene is the same gene as mouse genes, Mus

musculus chemokine MMRP2 mRNA of a GenBank accession

number U15209, Mus musculus C10-like chemokine mRNA of U19482 and mouse macrophage inflammatory protein-lgamma mRNA of U49513.

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 12. Only the amino acid sequence is shown in SEQ ID NO: 13.

#### Gene SCR-4

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The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 14. Only the amino acid sequence is shown in SEQ ID NO: 15.

It has been found that the sequence has a high homology to *Homo sapiens* clone 25077 mRNA of a GenBank accession number AF131820, and that it is considered to be a mouse ortholog. This sequence is described in WO 00/66784.

The nuclotide sequence of the gene from human and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 16. Only the amino acid sequence is shown in SEQ ID NO: 17.

### Gene SCR-5

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 18. Only the amino

acid sequence is shown in SEQ ID NO: 19.

It has been found that the sequence has a high homology with Homo sapiens esophageal cancer related gene 4 portein (ECRG4) mRNA of a GenBank accession number AF325503, and that it is considered to be a mouse ortholog of AF325503.

The nuclotide sequence of the gene from human and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 20. Only the amino acid sequence is shown in SEQ ID NO: 21.

#### Gene SCR-6

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The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 22. Only the amino acid sequence is shown in SEQ ID NO: 23.

#### Gene SCR-7

The nuclotide sequence of the gene from mouse and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 24. Only the amino acid sequence is shown in SEQ ID NO: 25.

### Gene SCR-8

25 The gene is the same gene as Mus musculus mRNA for ADAM23 of a GenBank accession number AB009673.

The nuclotide sequence of the gene from mouse and

the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 26. Only the amino acid sequence is shown in SEQ ID NO: 27.

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The sequence has a high homology with a sequence described by JP 11155574-A and the sequence described by JP 11155574-A is considered to be a human ortholog.

The nuclotide sequence of the gene from human and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 28. Only the amino acid sequence is shown in SEQ ID NO: 29.

Polypeptides which are products of these genes have an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells. The expression that a polypeptide has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells means that proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is supported in the presence of the polypeptide or in the presence of stroma cells expressing the polypeptide.

Therefore, the present invention provides use of the polypeptides and DNAs encoding the polypeptides and novel polypeptides among the polypeptides and DNAs encoding the novel polypeptides.

A stem cell proliferation-supporting factor which is

a polypeptide encoded by the DNA can be produced by introducing the DNA into a suitable host to prepare a transformant cell, and allowing the DNA to be expressed in the transformant cell.

The DNA may encode the above described factors which have amino acid sequences including substitution, deletion or insertion of one or several amino acids, as long as the activity of the stem cell proliferation—supporting factor to be encoded is not lost. DNAs encoding substantially equivalent polypeptides to this stem cell proliferation—supporting factor can be obtained by modifying the nucleotide sequences so as to include substitution, deletion, insertion, addition, or inversion of amino acid residues in a specific region using site—directed mutagenesis.

The DNAs including the above described mutation can be expressed in appropriate cells and the activity to support hematopoietic stem cells, of the expressed products can be examined, so that the DNAs encoding the polypeptide having functions which are substantially equivalent to this stem cell proliferation-supporting factor are obtained. In addition, the DNAs encoding substantially equivalently active protein as this stem cell proliferation-supporting factor can be obtained by isolating DNAs which hybridize with DNAs including, for example, the nucleotide sequence as described in SEQ ID NO: 8, 10, 12, 14, 16, 18, 20, 22, 24, 26 or 28 from the

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cells having the DNA, or probes prepared from these DNAs under the stringent condition; and which encode proteins possessing the activity to support hematopoietic stem cells. The length of the probe is usually 30 to 1000 nucleotides. The stringent condition is, for example, one in which DNAs having homology (determinable with homology search in the compare function of DNASIS version 3.7 (Hitachi Software Engineering)) at not less than 70%, preferably at not less than 80%, are hybridized each other and DNAs having less homology than those are not hybridized each other. The above described stringent condition may be 6 x SSC, 5 x

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pyrrolidone, 1% Ficoll 400) or 6  $\times$  SSC, 5  $\times$  Deanhardt, 0.5% SDS, 50% Formamide, 42°C, or the like.

citrate) (50 x Denhardt; 1% BSA, 1% polyvinyl

Denhardt, 0.5% SDS, 68°C (SSC; 3 M NaCl, 0.3 M sodium

Microorganisms such as Escherichia coli and yeast,

culture cells derived from animals or plants, and the like are used for host cells for expressing the DNA.

20 Preferably, culture cells derived from mammals are used as the host cells. In the case that prokaryotic cells are used as the host cells, the expression is preferably performed in a condition in which a signal peptide region is replaced with a leader sequence suitable for the prokaryotic cells such as β-lactamase (bla), alkaline phosphatase (phoA), and outer membrane protein A (ompA) and the like, or in a form in which a

methionine residue is added to the N-terminal site of the mature protein.

The introduction of the DNA to the host cell can be carried out by, for example, incorporating the DNA into a vector suitable for the host in an expressible form, and introducing the resultant recombinant vector to the host cell.

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Examples of the culture cells derived from mammals include CHO cell, 293 cell, COS7 cell, and the like.

10 Gene expression regulatory sequence such as a promoter to express the DNA may be originated from the gene itself, or may be derived from other genes such as cytomegalovirus promoter and elongation factor 1 promoter and the like.

15 Examples of a vector for animal culture cells include plasmid vectors, retrovirus vectors, adenovirus vectors (Neering, S.J., Blood, 88: 1147, 1996), herpes virus vectors (Dilloo, D., Blood, 89: 119, 1997), HIV vectors, and the like.

In order to introduce the recombinant vector into culture cells, the conventional methods which are usually employed for transformation of culture cells such as calcium phosphate transfection, the liposome method, the DEAE dextran method, the electroporation method and the microinjection method are employed.

The polypeptides of the present invention also comprise polypeptides having amino acid sequences in

which one or several amino acids are substituted,
deleted or inserted in the amino acid sequence
represented in SEQ ID NO: 9, 11, 13, 15, 17, 19, 21, 23,
25, 27 or 29, and having activity to support
hematopoietic stem cells in addition to the polypeptides
having the amino acid sequence represented in SEQ ID NO:
9, 11, 13, 15, 17, 19, 21, 23, 25, 27 or 29. That is,
even if mouse and human stem cell proliferation—
supporting factors are modified by substitution,
deletion, insertion or the like, polypeptides holding
essential functions as a stem cell proliferation—
supporting factor can be considered to be substantially
equivalent to the stem cell proliferation—supporting

factor.

These modified stem cell proliferation-supporting 15 factors can be obtained by treating DNA encoding the stem cell proliferation-supporting factor or host cells including the above mentioned DNA with a mutagen, or by mutating the above mentioned DNA so as to substitute, delete, or insert an amino acid at a specific site using 20 site-directed mutagenesis. The residual of the activity to support the hematopoietic stem cells in the obtained mutant polypeptide is confirmed by tranferring hematopoietic stem cells cultured in the presence of the mutant polypeptides into irradiated mice, and monitering 25 peripheral hematological cellularity over time, as in the examples described below.

As for the amino acid deletion, the polypeptide may be a fragment which lacks an amino acid sequence at the N-terminal end and/or the C-terminal end. The fragment lacking the amino acid sequence at the N-terminal end 5 and/or the C-terminal end can be obtained by a usual method, and the hematopoietic stem cell-supporting activity of the fragment can be determined by a similar way to that described with respect to the mutated polypeptide. In particular, if there is a portion predicted as a signal sequence or a transmembrane region 10 in the amino acid sequence, a fragment having the hematopoietic stem cell-supporting activity is predicted by using it as an index. For example, a protein encoded by human SCR-8 is a transmembrane protein of type I passing through the membrane once, and it is therefore 15 predicted that even if it made to be a soluble protein lacking the transmembrane region, it has the activity to support to proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells. The transmembrane region can be predicted with a known 20 program based on the amino acid sequence. For example, if it is predicted with a program called PSORT II (available through the Internet, URL: http://psort.nibb.ac.jp/index.html), the transmembrane region is amino acids at positions 790 to 806 in SEQ ID 25 NO: 29, and it is predicted that even if a fragment up to position 789, the fragment has activity to support

proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.

Since the nucleotide sequences of the above described DNAs have been clarified by the present invention, the DNAs can be also obtained by isolating the corresponding DNAs from mouse or human cDNA or chromosome DNA libraries using PCR in which the oligonucleotides prepared based on these nucleotide sequences are used as primers or using hybridization in which the oligonucleotides prepared based on these nucleotide sequences are used as probes.

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In order to complete the present invention, the DNAs of the present invention have been isolated from cDNA library of AGM-s3-A9 cells which are a mouse stromal cell line having the activity to support the hematopoietic stem cells, using SBH (Sequencing By Hybridization) method (Drmanac, S., Nat. Biotechnol., 16. 54, 1998; Drmanac, R., Methods. Enzymol., 303, 165, 1999) as described below. The mouse stromal cell lines having the activity to support the hematopoietic stem cells can be obtained using the method disclosed in W099/03980 or from Cell Bank of Institute of Physical and Chemical Research (RIKEN) or ATCC.

An outline of SBH method will be described below.

25 Probes having eight or nine nucleotides whose sequences are different from each other are prepared. When the nucleotide sequences corresponding to those of the probe

exist in a targeted gene, the probes can hybridize with the gene. The hybridization can be easily detected with utilization of radio isotope- or fluorescence-labelled probes. Each clone in the library is picked up, and 5 blotted on a membrane. Then, the repeated hybridizations are performed with the each of above described probes, so that one can identify the combination of probes that hybridize to each clone. Since the combination of hybridized probes depends on 10 genes, the combination of probes which hybridize to an identical gene is the same. That is, the same gene can be identified as one group (cluster) according to the the combination of the hybridized probes. The number of clones of each gene in the cDNA library can be determined by classifying each clone in the library 15 based on patterns of the hybridized probes and counting the classified clones. Thus, frequency of expression of each gene in the library can be determined.

activity to support the hematopoietic stem cells and from cells not having the activity. Clustering is performed for the cDNA libraries. Statuses of expressed genes among cells are compared, so that the genes highly expressed with specificity to the supporting cells are selected. The expression statuses of these genes in each of above described cells are further examined by Northern blot analysis, so that genes which are highly

expressed in the cells having the activity to support the hematopoietic stem cells are obtained.

The above mentioned genes are the genes which are highly expressed with specificity to the supporting cells and which are obtained through the above described process. Full-length genes have been cloned from the cDNA library derived from AGM-s3-A9 cell.

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Further, in order to determine an ability of gene products to support hematopoiesis, a gene fragment 10 including gene ORF was transferred into stromal cells using a retrovirus vector, and the change in the activity to support the hematopoietic stem cells of the stromal cells was determined. Specifically, after the stromal cells into which the gene was not introduced or 15 into which a control vector was introduced and those into which the gene was introduced were each co-cultured with the mouse hematopoietic stem cells, the hematopoietic cells were transplanted into irradiated mice. The engraftment of the co-cultured hematopoietic cells in recipient mice were examined. As a result, the 20 mice into which the hematopoietic stem cells co-cultured with the gene-introduced cells were transplanted, showed increased chimerism after the transplantation compared with co-culture with the cells into which the gene was 25 not introduced. This result shows that in the geneexpressed stromal cells, an activity to support the proliferation or survival of the hematopoietic stem

cells or the hematopoietic progenitor cells is increased or imparted. As a result, it has become evident that expression of the above described genes has a function to increase the above described activity in the stromal cells or impart the activity to the stromal cells. Therefore, it is revealed that products of the genes affect hematopoietic stem cells or hematopoietic progenitor cells to show an activity to support the survival or the proliferation thereof, or affect stromal cells to show an activity to increase an activity to support the hematopoietic stem cells therein or impart the activity thereto.

The polypeptides of the present invention can be used as a medicine to proliferate or support human 15 hematopoietic stem cells or human hematopoietic progenitor cells when they affect hematopoietic stem cells or hematopoietic progenitor cells to show an activity to support survival or proliferation thereof, in other words, the polypeptides have an activity to support survival or proliferation of hematopoietic stem 20 cells or hematopoietic progenitor cells if the hematopoietic stem cells or the hematopoietic progenitor cells are cultured in the presence of the polypeptides. The pharmaceutical composition can be used for 25 supporting proliferation or survival of human hematopoietic stem cells or human hematopoietic progenitor cells in vitro. For hematopoietic stem cell

transplantation therapies such as peripheral blood stem cell transplantation and cord blood stem cell transplantation, a sufficient amount of the hematopoietic stem cells sometimes cannot be collected and the transplantation may not be performed. Even if 5 the enough amount of the stem cells can not be collected, the enough amount of the hematopoietic stem cells can be obtained and transplanted by amplification of the hematopoietic stem cells in vitro using this polypeptides. That is, the hematopoietic stem cells can 10 be amplified without differentiation by culturing the hematopoietic stem cells in culture medium including these polypeptides. It may be considered the hematopoietic stem cells are able to be amplified more efficiently with addition of a variety of cytokines to 15 the medium.

when the hematopoietic stem cells or the hematopoietic progenitor cells are cultured in the medium including the polypeptides of the present invention, the hematopoietic stem cells or the hematopoietic progenitor cells used may be isolated one of these cell types alone or may be both of the cell types. In addition, the cells may include at least the hematopoietic stem cells or the hematopoietic progenitor cells, and include other hematopoietic cells. Further, it can be used a fraction containing hematopoietic stem cells or progenitor cells fractionated from the cell

population that contain the hematopoietic stem cells or progenitor cells.

Examples of sources of the hematopoietic stem cells and the hematopoietic progenitor cells in the method of the present invention include a fetal liver, bone marrow, fetal bone marrow, peripheral blood, the peripheral blood from persons whose stem cells are mobilized by administration of cytokines and/or antitumor drugs, cord blood, and the like of mammals such as human and mouse and the like. Any sources may be used as long as the tissue includes the hematopoietic stem cells.

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A culture method using petri dishes and flasks for culture may be employed to culture the hematopoietic stem cells or the hematopoietic progenitor cells. The cultivation of the hematopoietic stem cells and/or progenitor cells may be improved by mechanically controlling medium composition, pH, and the like, and using a bioreactor capable of high density cultivation (Schwartz, Proc. Natl. Acad. Sci. U.S.A., 88: 6760, 1991; Koller, M.R., Bio/Technology, 11: 358, 1993; Koller, M.R., Blood, 82: 378, 1993; Palsson, B.O., Bio/Technology, 11: 368, 1993).

The stromal cells in which DNAs encoding the polypeptide of the present invention can be obtained as described with respect to the expression of the DNAs.

The co-culture of the stromal cells and the hematopoietic cells can be performed simply after the

collection of the bone marrow cells without further separation. Furthermore, co-culture can be performed by separating components such as stromal cells, hematopoietic cells and other cell populations from collected bone marrow and combining them with the 3 hematopoietic cells and stromal cells which are not from the individual from which the bone marrow is cllected. Furthermore, after stromal cells are cultured to grow to the stromal cells, hematopoietic cells can be added to perform co-culture with the hematopoietic stem cells. 10 At this time, cell stimulating factors can added to the culture system of stromal cells to more effectively support proliferation and survival. Concrete examples of the cell stimulating factor include a growth factor 15 which is typically a cytokine such as SCF (stem cell factor), IL-3 (interleukin 3), GM-CSF (granulocyte/macrophage colony-stimulating factor), IL-6 (interleukin 6), TPO (thrombopoietin), G-CSF (granulocyte colony-stimulating factor), TGF-b 20 (transforming growth factor-b), MIP-la (Davatelis, G., J. Exp. Med. 167: 1939, 1988); a differentiation and proliferation control factor such as hematopoietic hormones such as EPO (erythropoietin), chemokine, Wnt gene product, and notch ligand; and a development 25 control factor.

In addition, when the polypeptide of the present invention affects hematopoietic stem cells or

hematopoietic progenitor cells to show an activity to support survival or proliferation thereof, in other words, the polypeptide has an activity to support survival or proliferation of hematopoietic stem cells or 5 hematopoietic progenitor cells if the hematopoietic stem cells or the hematopoietic progenitor cells are cultured in the presence of the polypeptide, the proliferation and the survival of the hematopoietic stem cells or the hematopoietic progenitor cells can be retained by allowing the recombinant polypeptide of the present 10 invention alone or in combination with the cell stimulating factors to affect hematopoietic stem cells or hematopoietic progenitor cells, without stromal cells. Examples of the cell stimulating factors used in this 15 case are above described cell stimulating factors and the like.

Medium used for the culture is not specially restricted as long as the proliferation or the survival of the hematopoietic stem cells or the hematopoietic progenitor cells is not harmed. Preferable media are, for example, MEM-α medium (GIBCO BRL), SF-02 medium (Sanko Junyaku), Opti-MEM medium (GIBCO BRL), IMDM medium (GIBCO BRL), and PRMI1640 medium (GIBCO BRL). A culture temperature is usually ranging from 25 to 39°C, and preferably ranging from 33 to 39°C. Examples of additives to the medium are fetal bovine serum, human serum, horse serum, insulin, transferrin, lactoferrin,

ethanolamine, sodium selenite, monothiolglycerol, 2mercaptoethanol, bovine serum albumin, sodium pyruvate,
polyethylene glycol, a variety of vitamins, and a
variety of amino acids. A concentration of CO<sub>2</sub> is
usually ranging from four to six percent, and preferably
five percent.

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Since the hematopoietic stem cells can differentiate into all the hematopoietic cell lineages, the hematopoietic stem cells can be amplified and

10 differentiated into a specific cell type in vitro, and then the specific cells can be transplanted. For example, when erythrocytes are necessary, after the cultivation of the patient's stem cells to amplify them, the hematopoietic cells whose main component is the erythrocyte can be artificially produced using an erythrocyte differentiation induction-promoting factor such as EPO.

The hematopoietic stem cells or the hematopoietic progenitor cells cultured using the polypeptides of the 20 present invention can be used as a graft for blood cell transplantation replacing the conventional bone marrow transplantation or cord blood transplantation.

Transplantation of the hematopoietic stem cells is superior to the conventional blood cell transplantation therapy, since the engraftment can last semipermanently.

The transplantation of the hematopoietic stem cells can be employed as therapy for a variety of diseases in

addition to combination therapy with total body X-ray irradiation therapy or advanced chemotherapy for leukemia. For example, when therapy accompanied with myelosuppression as an adverse reaction, such as 5 chemotherapy, radiation therapy, and the like is performed for the patient with solid cancer, the patient can get benefit of early recovery and stronger chemotherapy than the conventional one can be performed to improve the therapeutic effect of the chemotherapy by 10 collecting the bone marrow before the therapy, allowing the hematopoietic stem cells or the hematopoietic progenitor cells to be amplified in vitro and returning the amplified cells to the patient after the therapy. In addition, by allowing the hematopoietic stem cells or the hematopoietic progenitor cells obtained according to 15 the present invention to be differentiated into a variety of hematopoietic cells and transplanting these cells into a patient with hypoplasia of a given hematopoietic cells, the patient's deficient status can 20 be improved. In addition, this therapy can improve dyshemopoietic anemia to develop anemia such as aplastic anemia caused by bone marrow hypoplasia. Furthermore, examples of diseases in which the transplantation of the hematopoietic stem cells according to the present invention is effective include immunodeficiency syndrome 25 such as chronic granulomatous disease, duplicated immunodeficiency syndrome, agammaglobulinemia, WiskottAldrich syndrome, acquired immunodeficiency syndrome (AIDS), and the like, thalassemia, hemolytic anemia due to an enzyme defect, congenital anemia such as sicklemia, Gaucher's disease, lysosomal storage disease such as mucopolysaccharidosis, adrenoleukodegeneracy, a variety of cancers and tumors, and the like.

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Transplantation of the hematopoietic stem cells may be performed in the same manner as the conventional bone marrow transplantation or cord blood transplantation other than the differences of the cells used.

The source of the hematopoietic stem cells which may be used for the above described hematopoietic stem cell transplantation is not restricted to the bone marrow, and the above described fetal liver, the fetal bone marrow, the peripheral blood, the peripheral blood with stem cells mobilized by administration of cytokines and/or antitumor drugs, the cord blood, and the like may be used.

The graft may be a composition including buffer solution and the like in addition to the hematopoietic stem cells and the hematopoietic progenitor cells produced by the method according to the present invention.

The hematopoietic stem cells or the hematopoietic

25 progenitor cells produced according to the present
invention may be used for ex vivo gene therapy. Because
of the low frequency of recombination of target genes to

the chromosome because the stem cells are in the resting state, differentiation of the stem cells during the culture period, and the like, the gene therapy to the hematopoietic stem cells has been hard to be established. However, the present invention can amplify the stem 5 cells without differentiation, so that efficacy of gene transfer is expected to be remarkably improved. In this gene therapy, a foreign gene (a gene for therapy) is transferred into the hematopoietic stem cells or the hematopoietic progenitor cells, and then the obtained 10 gene-transferred cells are used. The foreign gene to be transferred is appropriately selected according to disease. Examples of diseases in which the target cells of the gene therapy are the hematopoietic cells include immunodeficiency syndrome such as chronic granulomatous 15 disease, duplicated immunodeficiency syndrome, agammaglobulinemia, Wiskott-Aldrich syndrome, acquired immunodeficiency syndrome (AIDS), and the like, thalassemia, hemolytic anemia due to an enzyme defect, congenital anemia such as sicklemia, Gaucher's disease, 20 lysosomal storage disease such as mucopolysaccharidosis, adrenoleukodegeneracy, a variety of cancers and tumors,

A usual method used for transfer of a gene into

25 animal cells is employed for the transfer of the gene
for the therapy into the hematopoietic stem cells or the
hematopoietic progenitor cells. Examples include a

and the like.

method using a vector for animal cells derived from virus utilized for a gene therapy such as retrovirus vectors such as Moloney mouse leukemia virus, adenovirus vectors, adeno-associated virus (AAV) vectors, herpes 5 simplex virus vectors, and HIV vectors (with respect to a vector for gene therapy, see Verma, I.M., Nature, 389: 239, 1997); calcium phosphate transfection, DEAE-dextran transfection, electroporation, the liposome method, the lipofection method, the microinjection method, and the 10 like. Among them, the method using the retrovirus vector, the adeno-associated virus vector, or the HIV vector is preferable, since permanent expression of a gene is expected due to insertion into the chromosome DNA of a target cell.

15 For example, the adeno-associated virus (AAV) vector can be prepared as follows. First, a vector plasmid in which a gene for therapy is inserted into ITR (inverted terminal repeat) at both ends of wild-type adenoassociated virus DNA and a helper plasmid for 20 supplementing virus proteins are transfected into 293 cell line. Next, adenovirus as helper virus is infected, so that virus particles including the AAV vector are produced. Alternatively, instead of adenovirus, a plasmid which expresses adenovirus gene having helper 25 function may be transfected. The hematopoietic stem cells or the hematopoietic progenitor cells are infected with the obtained virus particles. Preferably,

appropriate promoter, enhancer, insulator and the like are inserted into the upstream region of the target gene in the vector DNA, so that the expression of the gene is regulated. When a marker gene such as a drug resistant gene is used in addition to the gene for therapy, cells into which the gene for therapy are transferred are easily selected. The gene for therapy may be a sense gene or an antisense gene.

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A composition for gene therapy may include a buffer solution and a novel active ingredient and the like in addition to the hematopoietic stem cells or the hematopoietic progenitor cells by the method according to the present invention.

A vector for gene therapy can be produced by incorporating the DNA of the present invention in an expression vector using a usual method. This vector for gene therapy is useful to treat diseases which need survival and proliferation of the human hematopoietic stem cells. That is, the vector for gene therapy is transferred into the hematopoietic stem cells and the cells are cultured in vitro, so that the hematopoietic stem cells or the hematopoietic progenitor cells can proliferate dominatingly. The proliferation of hematopoietic stem cells in vivo can be caused by 25 returning these hematopoietic stem cells thus treated. The proliferation of hematopoietic stem cells in vivo can significantly promoted by introducing this vector

for gene therapy into the body. Alternatively, the bone marrow cells derived from a patient are cultured as it is and this vector for gene therapy is transferred thereto, so that the hematopoietic stem cells or the 5 hematopoietic progenitor cells can be proliferated in a culture system. Alternatively, this vector for gene therapy is transferred into the stromal cells and mesenchaymal stem cells obtained by isolating and culturing stromal cells from the bone marrow, so that 10 the activity to support the hematopoietic stem cells can be added or increased.

As shown in Examples, since it is possible that by introducing the DNA of the present invention into the stromal cells without the activity to support the hematopoietic stem cells, this activity can be imparted, stromal cells having the activity to support the hematopoietic stem cells can be prepared by gene transfer to stromal cells derived from human or mouse. The stromal cells expressing the DNA of the present 20 invention and the hematopoietic stem cells or the hematopoietic progenitor cells are co-cultured, so that the hematopoietic stem cells or the hematopoietic progenitor cells can survive and proliferate so as to be useful for a variety treatment.

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Since the hematopoietic stem cells or the 25 hematopoietic progenitor cells can survive and proliferate by expression of the DNA of the present invention in the stromal cell, an activity to support
the hematopoietic stem cells of the stromal cells can be
determined using the expression of the DNA of the
present invention as an index. The expression of the

5 DNA of the present invention in the stromal cells can be
confirmed using an antibody against a polypeptide
encoded by the DNA of the present invention. Also, PCR
primers can be prepared based on nucleotide sequences,
and RNA is prepared from the stromal cells of interest,

10 and RT-PCR is performed, so that the expression of the
DNA of the present invention can be confirmed. The
antibody will be described below.

The pharmaceutical composition of the present invention can be administered to human. 15 pharmaceutical composition having an activity to proliferate or to support the human hematopoietic stem cells or the hematopoietic progenitor cells can be produced by mixing medically acceptable diluent, stabilizer, carrier, and/or other additives with the polypeptides of the present invention. At this time, in 20 order to increase the stability of the protein in vivo, the polypeptides of the present invention may be modified by a modifying agent. Examples of the modifying agent include polyethylene glycol (PEG), dextran, poly(N-vinyl-pyrrolidone), polypropylene glycol 25 homopolymer, polypropylene oxide/ethylene oxide copolymer, polyoxyethylated polyol, polyvinyl alcohol,

and the like. The modification of the protein with PEG can be performed by, for example, a method in which activated ester derivatives of PEG is reacted with the protein, a method in which aldehyde derivatives at the terminal portion of PEG is reacted with the protein in the presence of a reducing agent, and the like.

Japanese Patent Application Laid-Open No. 10-510980 discloses such protein modification in detail.

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invention is administered to human, recovery from hematological suppression due to an adverse drug reaction of carcinostatics; early recovery of hematopoietic cells at bone marrow transplantation, peripheral blood stem cell transplantation, or cord blood transplantation; and recovery of hematopoietic function at pancytopenia such as aplastic anemia (AA) and myelodysplastic syndrome (MDS) are expected.

The antibodies of the present invention react specifically to the above described polypeptides of the present invention. The antibodies of the present invention may be monoclonal antibodies or polyclonal antibodies as long as they react specifically to the above described polypeptides.

The antibodies of the present invention can be

25 prepared according to usual methods. For example, the
antibodies can be prepared either in vivo method in
which animals are additionally immunized by an antigen

together with adjuvant once or several times at an interval of several weeks or in vitro method in which immune cells are isolated and sensitized in an appropriate culture system. Examples of immune cells which can produce the antibodies of the present invention include splenic cells, tonsillar cells, lymph gland cells, and the like.

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The whole polypeptide according to the present invention is not necessarily used as an antigen. A part of this polypeptide may be used as an antigen. When the 10 antigen is a short peptide, particularly, a peptide made of about 20 amino acid residues, it may be used by binding it to a carrier protein having high antigenicity such as keyhole lympet hemocyanin or bovine serum 15 albumin using chemical modification and the like. Alternatively, the antigen may be used by covalently binding it to peptide having branching skeleton such as lysine core MAP peptide instead of the carrier protein (Posnett et al., J. Biol. Chem., 263, 1719-1725, 1988; Lu et al., Mol. Immunol., 28, 623-630, 1991; Briand et 20 al., J. Immunol. Methods, 156, 255-265, 1992).

Examples of adjuvant include Freund's complete adjuvant, Freund's incomplete adjuvant, aluminum hydroxide gel, and the like. Antigen-given animals are, for example, mouse, rat, rabbit, sheep, goat, chicken, bovine, horse, guinea pig, hamster, and the like. The blood is collected from these animals and the serum is

separated. Then, immunoglobulin is purified from the serum using an ammonium sulfate precipitation method, anion exchange chromatography, protein A chromatography, or protein G chromatography to obtain polyclonal antibodies.

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With respect to chicken, antibodies can be purified from an egg. Monoclonal antibodies can be prepared by purification from supernatant of culture of hybridoma cells which are made by fusion of the immune cells 10 sensitized in vitro, or immune cells from the above described animals with parent cells capable of cultivation, or ascites from animals which received intraperitoneal administration of hybridoma cells. Examples of parent cells include X63, NS-1, P3U1, X63.653, SP2/O, Y3, SK0-007, GM1500, UC729-6, HM2.0, 15 NP4-1 cell lines, and the like. Preparation may be performed by cultivating the immortalized antibodyforming cells obtained by sensitization in vitro, or infection of a proper virus such as EB virus to the 20 immune cells of the above described animals.

In addition to these cell engineering methods, the antibodies can be obtained using gene engineering methods. For example, the antibody gene obtained from the *in vitro* sensitized cells or immune cells derived from the above described animals is amplified by PCR (polymerase chain reaction) and isolated, and the amplified genes are transferred into microorganisms such

as *E. coli* to produce the antibodies. Alternatively, the antibodies may be expressed on surfaces of phages as fused proteins.

By measuring polypeptides in vivo using the antibodies of the present invention, the relationship between the polypeptides and pathological status of a variety of diseases can be clarified. Moreover, the antibodies can be used for diagnosis and treatment of diseases, and efficient affinity purification of the polypeptides.

The present invention provides polypeptides having an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells by effecting thereon, or an activity to impart an activity to support the hematopoietic stem cells to stromal cells by effecting thereon, and also provides DNAs encoding thereof. The polypeptides of the present invention can efficiently maintain the proliferation or the survival of the hematopoietic stem cells or the hematopoietic progenitor cells.

# Best Mode for Carrying out the Invention Hereafter, the present invention will be described in detail by reference to examples.

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Example 1 Preparation of fragment of gene which is specifically expressed in hematopoietic stem cell-

### supporting cells

- (I) Preparation of stromal cell line derived from mouse AGM
- (1) Isolation of AGM region from fetal mouse 2 C3H/HeNSLc mice of both genders (purchased from Japan SLC INC.) were kept under a SPF (specific pathogen-free) environment. One or two female mice and one male mouse were placed in the same cage over a night. In the next morning, the female mice in which the existence of a vaginal plug was observed were 10 transferred to other cages and kept. The day when the existence of the vaginal plug was observed was defined to be the 0.5th day of pregnancy. On the 10.5th day of the pregnancy, after mouse was sacrificed by cervical dislocation, fetuses were extirpated. Isolation of AGM 15 regions was performed according to the method by Godin et al. (Godin, I., Proc. Natl. Acad. Sci. U.S.A., 92: 773-777, 1995) and the method by Medvinsky et al. (Medvinsky, A.L., Blood, 87: 557-565, 1996). The 20 fetuses were placed in a culture dishes to which PBS(-) (phosphate buffered saline) (produced by Nissui Seiyaku) was added in a volume just sufficient to cover the fetuses. After the AGM regions were carefully excised so as not to include other regions under a stereoscopic microscope, they were put in another 24-well culture 25 dish (Nunc).
  - (2) Establishment of cell lines derived from AGM

One drop of MEM medium (Sigma) containing 10% FCS (Hyclone) was added to the AGM regions in the 24-well culture dish (Nunc), and AGM regions were cultured in an incubator overnight. The culture was performed in the MEM medium (Sigma) containing 10% FCS (Hyclone) at 37°C, in an atmosphere of 5% CO<sub>2</sub>, and at a humidity of 100%. When the cells of the AGM regions adhered to the culture dish due to overnight cultivation, two milliliters of MEM medium containing 10% FCS was further added.

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- 10 Stromal cells began to appear around the AGM region tissue fragment after the continuous cultivation. After one-week cultivation, adhesive cells were separated by trypsin treatment (0.05% trypsin in PBS containing 0.53 mm EDTA (Gibco BRL) at 37°C for three to five minutes).
- The stromal cells were then washed twice with the medium, and seeded on 6-well culture dish (Nunc). On the next day, the cells which did not adhere to the culture dish and the medium were removed, and then, fresh medium was added. Two weeks after transfer to the 6-well culture
- 20 dish, cells were γ-ray-irradiated at 900 Rad to eliminate endogenous hematopoietic cells. An attempt of the direct cell cloning by limiting dilution from this culture system was made, but no cell proliferation was observed and the cloning ended in failure. Then, after
- 25 the number of seeded cells in one well was increased and cells were adapted so as to be able to proliferate from a small number of cells, the cells were cloned by

limiting dilution.

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bone marrow

Specifically, the AGM was extirpated and cultured in the same manner as described above. The culture system two weeks after the γ-ray radiation was trypsinized

5 (0.05% trypsin in PBS containing 0.53 mM EDTA at 37°C for three to five minutes) to suspend the cells, and the cells were seeded in a 24-well culture dish at 50 to 100 cells/well. After the culture was continued for three weeks, the cells were seeded in a 96-well culture dish

10 (Nunc) by means of limiting dilution, at 0.3 cells/well. The cells which were grown from the well in which only one cell was seeded were allowed to enlarge culture. As a result, the cells were successfully cloned to obtain fibroblast-like cells and cobble stone-like cells.

A CD34-positive cell fraction derived from the human cord blood was co-cultured with the fibroblast-like cells for two weeks to examine the presence of colony-forming cells during the culture. Colony-forming cells could not be found in the co-culture system with the fibroblast-like cells. Then, the similar examination was performed for seven cell clones showing the cobblestone-like form. Three clones having an activity to support proliferation of human hematopoietic stem cells were obtained and were named AGM-s1, AGM-s2, and AGM-s3.

Bone marrow was collected from a femur of C57BL/6-

Ly5.1 pep (eight- to ten-week age, and male) (the gift from Professor K. Nakauchi, University of Tsukuba), and suspended in PBS. After the mouse bone marrow mononuclear cells were concentrated by specific gravity centrifugation according to the usual method (S. Kouzu, Fundamental techniques for immunology, YODOSHA, 1995), the cells were suspended in staining buffer (PBS containing 5% FCS and 0.05% NaN3), and a hematopoietic stem cell fraction was obtained as follows (Osawa, M. et al., Science 273: 242-245, 1996).

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An FITC-conjugated anti-CD34 antibody, a phycoerythrin-conjugated anti-Sca-1 antibody, an allophycocyanin anti-c-Kit antibody (all purchased from Pharmingen) and six biotylated anti-differentiation antigen antibodies (CD45R, CD4, CD8, Gr-1, Ter119, and 15 CD11c, all purchased from Pharmingen) as molecular markers (Lin), were added to a suspension of the bone marrow mononuclear cells and incubated for 20 min on ice to cause reaction. After the cells were washed twice with staining buffer, CD34-negative, Sca-1-positive, c-20 Kit-positive, and Lin-negative cells were isolated on a cell sorter (FACS Vantage, Becton Dickinson). (III) Subcloning of mouse stromal cell line and determination of activity to support hematopoietic stem cells of a variety of cell lines 25

- (1) Subcloning of mouse stromal cell line
- 1) Isolation of AGM-s3 subclone

Stromal cell line AGM-s3 derived from AGM, which was subcultured in MEMa medium (GIBCO BRL) including inactivated 10% FCS (bovine fetal serum, Hyclone), was suspended in PBS containing 5% FCS (PBS-FCS). Clone sorting was performed in a 96-well culture dish (Falcon) at one cell/well using a cell sorter (FACS Vantage; Becton Dickinson). Among cells in the 96 wells, cultures of the cells which grew were expanded, so that thirteen kinds of AGM-s3 subclones were obtained. The activity to support the hematopoietic cells of these AGM-s3 subclones were examined.

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2) Isolation of human cord blood CD34-positive stem cell The human cord blood was collected at normal delivery according to the criteria approved by Ethics 15 committee of Kirin Beer Iyaku Tansaku Kenkyusho. cord blood was collected using a heparin-added syringe so as not to coaqulate. The heparin treated cord blood was overlaid on Lymphoprep (NYCOMED PHARMA), and mononuclear cells were separated by specific gravity 20 centrifugation (at 400G, at room temperature, and for 30 minutes). Erythrocytes contaminated in the mononuclear cell fraction were lyzed by treatment with an ammonium chloride buffer solution (0.83% NH4Cl-Tris HCl, 20 mM, pH 6.8) at room temperature for two minutes. After the 25 mononuclear cells were washed with PBS-FCS, ten milligrams of human IgG was added thereto and the mixture was allowed to stand on ice for ten minutes.

Then, the cells were further washed with PBS-FCS.

Biotinylated antibodies against the antigens specific to the human differentiated blood cells, that is, the antibodies against CD2, CD11c (purified from ATCC

- 5 hybridoma), CD19 (Pharmingen), CD15, and CD41 (Leinco Technologies Inc.), and Glycophorin A (Cosmo Bio) were added thereto, and the mixture was allowed to stand on ice for 20 min. After washing with PBS-FCS, the cells were suspended in one milliliter of PBS containing 5%
- 10 FCS, 10 mM EDTA, and 0.05% NaN3 (PBS-FCS-EDTA-NaN3).

  Streptavidin-bound magnetic beads (BioMag. Per Septive Diagnostics) were added thereto, and the mixture was allowed to stand on ice for 40 min. The differentiated blood cells which expressed differentiation antigens
- Dynal). An FITC-labeled anti-CD34 antibody (Immunotech S.A., Marseilles, France) was added to the remaining differentiated blood cell antigen-negative cell fraction.

  After incubation on ice for 20 min., a CD34-positive
- 20 fraction was recovered using a cell sorter. This cell population was defined as a hematopoietic stem cell population derived from the human cord blood.
  - 3) Co-culture of the human hematopoietic stem cells and AGM-s3 subclone
- 25 After 13 kinds of AGM-s3 subclones and stromal cell line MS-5 derived from the mouse bone marrow were each seeded in a 24-well culture dish (Falcon) at  $1\times10^4$

cells/well, and cells were cultured in one milliliter of MEMα medium containing 10% FCS and allowed to grow until the cells covered all over the bottom surfaces of the wells. CD34-positive hematopoietic stem cells derived from the human cord blood were placed on the above 5 described stromal cells at 500 cells/well, and cocultured in one milliliter of MEMa medium containing 10% FCS. One week after the start of the co-culture, one milliliter of the same medium was further added. Two weeks after the start of the co-culture, the stromal 10 cells and the human blood cells were trypsinized (0.05% trypsin in PBS containing 0.5 mM EDTA (GIBCO BRL) at 37°C; standing for two to five min.) to simultaneously separate them from the culture dish. An activity to 15 support the hematopoietic stem cells was determined with a clonogenic assay.

- 4) Assessment of proliferation statuses of the hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay
- The cells which proliferated in the above described co-culture system were appropriately diluted, and subjected to one milliliter of methylcellulose culture system to be analyzed. The analysis using the methylcellulose culture system was performed using a 6-well culture dish (Falcon) in MethoCult H4230 (Stem Cell Technologies Inc.) to which 10 ng/ml of human SCF, human IL-3, human IL-6, human G-CSF, and human TPO, and 2

IU/ml of EPO were added. All of a variety of the above described hematopoietic factors were recombinants and pure. After two-week culture, developed colonies were observed under a microscope to count numbers of CFU-GM (granulocyte-macrophage colony-forming unit), BFU-E (erythroid burst forming unit), and CFU-E mix (erythrocyte mixed colony-forming unit).

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Fig. 1 shows the result of two-week co-culture of the CD34-positive hematopoietic stem cells and the AGM-10 s3 subclone A9, A7, or D11. As a result of the coculture, A9 and D11 subclones among 13 kinds of AGM-s3 subclones supported proliferation of all three series of CFU-GM, BFU-E, and CFU-E mix. Especially, although BFU-E and CFU-E mix, that is, the progenitor cells of erythrocytes were hardly to be supported in usual, their 15 proliferations were observed in the co-culture system with A9 or D11 cells. The results showed that proliferation or maintenance of the hematopoietic stem cells or the hematopoietic progenitor cells occurred in the co-culture with A9 or D11 cells and the progenitor 20 cells of the erythrocyte were continuously supplied. In contrast, although cellular morphology of A7 was similar to that of A9, A7 did not support CFU-GM, BFU-E, and CFU-E mix.

5) Comparison of an activity to support the human hematopoietic stem cells between A9 and a stromal cell line OP9 derived from mouse fetus

Comparison of an activity to support the CD34positive hematopoietic stem cells derived from the human cord blood between AGM-s3 subclones A9 and A7, and a stromal cell line OP9 derived from mouse fetus (RCB1124, the Cell Development Bank of RIKEN) were performed with 5 CFU-GM, BFU-E, CFU-E and CFU-E mix as indexes, using the above described determination system. Fig. 2 shows the result of the two-week co-culture. In the A7 cell culture system, CFU-GM, BFU-E, and CFU-E were significantly decreased and CFU-E mix was completely 10 disappeared. In contrast, with OP9 cells, a variety of blood cell progenitor cells including CFU-E mix were supported, although the supporting ability was less than that of A9 cells. Therefore, it has been found that OP9 cells possess the activity to support the hematopoietic 15 stem cells.

(2) Assessment of activity to support the hematopoietic stem cells in a variety of cell lines

The above described stromal cell lines (AGM-s3-A9,

20 AGM-s3-A7, and AGM-s3-G1), 3T3Swiss (ATCC), OP9, and

NIH3T3 (ATCC) were seeded in a 24-well culture dish

(Falcon) at 5 × 10<sup>4</sup> cells/well. The cell lines were

cultured in MEMα medium (GIBCO BRL) containing

inactivated 10% FCS (bovine fetal serum, Hyclone) for

25 one day and allowed to proliferate until the cells

covered all over the bottom surfaces of the wells. Then,

the medium was replaced to one milliliter of fresh

medium, thirty cells of the mouse hematopoietic stem cells (derived from C57BL/6-Ly5.1) obtained in the above (II) were placed on this cell layer, and co-culture was started.

On seventh day of the cultivation, the cells were 5 trypsinized (0.05% trypsin in PBS containing 0.5 mM EDTA (GIBCO BRL) at 37°C for two to five minutes) to separate and recover all the cells on the culture dish. recovered whole cells of each cell line and 200,000 cells of whole bone marrow cells (derived from C57BL/6-10 Ly5.2 mouse, Charles River) were transplanted into C57BL/6-Ly5.2 mice (eight weeks age and male, Charles River) irradiated with X-ray at 8.5 Gy through the tail vein. After the transplantation, peripheral blood was collected from orbit at intervals, and the ratio in 15 number of cells derived from the C57BL/6-Ly5.1 prep mouse was determined with FACS. The peripheral blood was analyzed according to the usual method (S. Kouzu, Fundamental techniques for immunology, YODOSHA, 1995). 20 Three hundreds and fifty µL of distilled water was added to 50  $\mu L$  of the peripheral blood, and the mixture was allowed to stand for 30 seconds so as to lyze the erythrocytes. Then, PBS at twice concentrations was added and the mixture was centrifuged to recover white 25 blood cells. After the cells were washed once using the staining buffer (PBS containing 5% FCS and 0.05% NaN3), anti-CD16 antibody, anti-Ly5.1 (CD45.1) antibody labeled with FITC, anti-Gr-1 and anti-CD11c antibodies labeled with phycoerythrin, and anti-CD45R (B220) and anti-CD90 (Thyl) antibodies labeled with allophycocyanin (all of these were purchased from Pharmingen) were added. After these cells were allowed to stand for reaction in the ice bath for 30 minites, they were washed with the staining buffer and FACS analysis was performed.

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Change in the number of cells capable of reconstitution during the hematopoietic stem cell culture was determined by calculating the proportions of Ly5.1-positive cells in the Gr-1- or CD11c-positive cells (myeloid cells) and Ly5.1-positive cells in the CD90- or CD45R-positive cells (lymphoid cells) in the peripheral blood at intervals after transplantation.

Fig. 3 shows the results. When the cells were co-cultured with AGM-s3-A9 cells, OP9 cells, or 3T3Swiss cells, high chimerism of donor cells were maintained after the transplantation. Therefore, these stromal cells were considered to have a high activity to support the hematopoietic stem cells. In contrast, when the cells were co-cultured with AGM-s3-A7 cells, AGM-s3-G1 cells, or NIH3T3 cells, high chimerism derived from the transplanted cells was not observed. Therefore, these stromal cells were low in an activity to support the hematopoietic stem cells or the hematopoietic progenitor cells.

(IV) Identification of sequences of genes which

specifically express in hematopoietic stem cellsupporting cells

AGM-s3-A9 cells, AGM-s3-A7 cells and OP9 cells were each dissolved in 20 mL of ISOGEN (Nippon gene, Japan) and total RNAs were prepared according to the attachment. 5 Messenger RNAs were prepared from one milligram of the total RNAs according to the protocol of the mRNA purification kit (Amersham Pharmacia, U.S.A.). cDNAs were synthesized from the mRNAs and cDNA libraries 10 (hereinafter, also called as AGM-s3-A9 cDNA, AGM-s3-A7 cDNA and OP9 cDNA, respectively) were constructed using pSPORT1 (GIBCO Lifetech, U.S.A.). A clone harboring a cDNA fragment which highly expresses specifically to AGM-s3-A9 cells or OP9 cells compared with AGM-s3-A7 cells was obtained from the libraries with SBH method 15 (Hyseq, U.S.A.). A nucleotide sequence of the obtained clone was determined using ABI377 DNA sequencer (Perkin Elmer, U.S.A.).

As a result, it has been found that expression of

genes comprising nucleotide sequences shown in SEQ ID

NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5,

SEQ ID NO:6, and SEQ ID NO:7, or parts thereof in AGM
s3-A9 or OP9 cells is higher than that in AGM-s3-A7

cells. These genes were named as SCR-2, SCR-3, SCR-4,

SCR-5, SCR-6, SCR-7 and SCR-8, respectively.

# Example 2 Cloning of SCR-2 and activity determination

By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 1 with BLAST, it has been found that SCR-2 is the same gene as a mouse gene, Mus musculus glypican-1 (Gpc-1) of an accession number AF185613. The nuclotide sequence of ORF (Open Reading Frame) of SCR-2 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 8. Only the amino acid sequence is shown in SEQ ID NO: 9.

The human nucleotide sequence of Gpc-1 is recorded in GenBank database under an accession number AX020122.

The nucleotide sequence of ORF of AX020122 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 10. Only the amino acid sequence is shown in SEQ ID NO: 11.

Determination of the activity to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of 20 mouse SCR-2

Based on the nucleotide sequence of SCR-2 ORF, SCR-2Fsall and SCR-2Reco primers having the following nucleotide sequences were prepared, and PCR was performed using OP9 cDNA as a template.

25 SCR-2Fsal

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CCGGTCGACCACCatggaactccggacccgaggctgg (SEQ ID NO: 30)

SCR-2Reco

CCGAATTCttaccgccacctgggcctggctgc (SEQ ID NO: 31)

An amplified fragment was digested with restriction enzymes EcoRI and SalI. After electrophoresis, a DNA fragment was purified using JETSORB (Genomed, Germany). 5 The purified DNA fragment was ligated with pMX-IRES-GFP vector digested with EcoRI and XhoI (gift form Professor T. Kitamura, TOKYO UNIV. INST. OF MEDICAL SCIENCE, Japan). The pMX-IRES-GFP vector is a plasmid obtained 10 by inserting sequences encoding IRES (Internal Ribosome Entry Site) and GFP (Green Fluorescence Protein) into the retrovirus vector pMX. IRES (Internal Ribosome Entry Site) enables ribosome to access to the middle of the mRNA. Therefore, two genes can be expressed from one mRNA by ligation of upward and downward genes 15 separated by IRES in one transcription unit during the construction of an expression vector. With respect to the above-described plasmid, SCR-2 cDNA was inserted in the upward site and GFP (Green Fluorescence Protein) was 20 inserted in the downward site. Thus, the expression of SCR-2 could be monitored by detecting the expression status of GFP using FACS.

The obtained recombinant vector was introduced into E. coli DH5α, and was seeded on LB agar medium

25 containing 100 μg/ml of ampicillin, so that independent colonies were formed. After the isolated colony was cultured in 100 mL of LB medium containing 100 μg/ml of

ampicillin, plasmid was purified using QIAGENtip100 (QIAGEN, U.S.A.). The sequence of the inserted gene was determined using a conventional method, so that the sequence was confirmed to be identical to the nucleotide sequence of SCR-2 ORF.

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(2) Preparation of stromal cells highly expressing SCR-2 BOSC23 cells were seeded on a collagen type I-coated 60-mm dish (Asahi technoqlass) at  $2 \times 10^6$  cells/dish, and cultured in DMEM medium containing 10% FCS at 37°C, under an atmosphere of 5% CO2, and at a humidity of 100%. Twelve to 18 hours after the start of the culture, the medium was replaced by two milliliters of OPTI MEM medium (GIBCO BRL).

About 3 µg of plasmid obtained by inserting SCR-2 into the above described pMX-IRES-GFP was added to 18  $\mu$ l of LIPOFECTAMINE Reagent (GIBCO BRL) diluted with 100 ul of OPTI MEM medium, and the mixture was allowed to stand at room temperature for 30 min. The prepared DNA solution was added to the prepared BOSC23 cell culture 20 solution. After about five hours, two milliliters of DMEM medium containing 20% FCS (GIBCO BRL) was added.

After about 24 hours, the medium was replaced by 4 ml of DMEM containing 10% FCS. Further, after about 48 hours, the culture medium was harvested. After the culture medium was filtrated through 0.45-µm filter, the filtrate was centrifuged at 1,200g for 16 hours and the supernatant was removed to obtain the virus

precipitation.

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AGM-s3-A7 or AGM-s3-A9 cells were cultured in one milliliter of MEMa medium containing 10% FCS (GIBCO BRL) on a 24-well culture dish (FALCON) at  $1 \times 10^4$  cells/well. After 12 to 18 hours, the virus precipitation was suspended in one milliliter of MEMa medium containing 10% FCS, and the stromal cell culture medium was replaced by the virus suspension. Next, POLYBRENE (Sigma, SEQUA-BRENE) was added to be 10 µq/ml. After the culture dish was centrifuged at 700g for 45 minutes, 10 the cells were cultured at 37°C, under an atmosphere of 5% CO2, and at a humidity of 100%. After 48 hours, the medium was replaced by one milliliter of MEMa medium containing 10% FCS. After 24 hours, the cells were subcultured on a 6-well culture dish (FALCON) and 15 cultured in three milliliters of MEMa medium containing 10% FCS. Forty-eight hours after the subculturing, GFP expression in the stromal cells was detected using a cell sorter (FACSVantage, Becton Dickinson) to indirectly confirm that not less than 80% of cells 20 expressed SCR-2.

Also, the same procedures were repeated by using pMX-IRES-GFP vector instead of the plasmid obtained by inserting SCR-2 into pMX-IRES-GFP to prepare stromal cells into which a control vector was introduced.

(3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-2, and determination

of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay

In the same manner as described in (III) (1) 3) to
4) of Example 1, AGM-s3-A9 or AGM-s3-A7 cells in which
5 SCR-2 was highly expressed through retrovirus, AGM-s3-A9
or AGM-s3-A7 cells into which a control vector was
introduced, or AGM-s3-A9 or AGM-s3-A7 cells were cocultured with CD34-positive hematopoietic stem cells
derived from human cord blood, and proliferation
10 statuses of hematopoietic stem cells and hematopoietic
progenitor cells are determined.

Fig. 4 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9 cells in which SCR-2 was highly expressed (A9/SCR-2), AGM-S3-A9 cells into which a control vector was 15 introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks. Also, Fig. 5 shows results when the CD34positive hematopoietic stem cells were co-cultured with AGM-S3-A7 cells in which SCR-2 was highly expressed, AGM-S3-A7 cells into which a control vector was 20 introduced or AGM-S3-A7 cells for two weeks. As a result, by the co-culture with AGM-S3-A9 cells in which SCR-2 was highly expressed or AGM-S3-A7 cells in which SCR-2 was highly expressed, increases of BFU-E and CFU-C 25 were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 or AGM-S3A7 increases by allowing SCR-2 to be highly expressed. From the results, it has been revealed that a gene product of SCR-2 has an activity to support survival or proliferation of hematopoietic stem cells or

hematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

# 10 Example 3 Cloning of SCR-3 and activity determination

By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 2 with BLAST, it has been found that SCR-3 is the same gene as mouse genes, Mus musculus chemokine MMRP2 mRNA of an accession number U15209, Mus musculus C10-like chemokine mRNA of U19482 and mouse macrophage inflammatory protein-lgamma mRNA of U49513. The nuclotide sequence of SCR-3 ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 12. Only the amino acid

20 sequence is shown in SEQ ID NO: 13.

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Determination of the activity of SCR-3 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of 25 mouse SCR-3

Based on the nucleotide sequence of SCR-3 ORF, SCR-3FxhoI and SCR-3Reco primers having the following

nucleotide sequences were prepared, and PCR was performed using AGM-s3-A9 cDNA as a template. An amplified fragment was inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in (1) of Example 2.

SCR-3FxhoI

ccgCTCGAGccaccATGAAGCCTTTTCATACTGCC (SEQ ID NO: 32)
SCR-3Reco

tccGAATTCttattgtttgtaggtccgtgg (SEQ ID NO: 33)

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- (2) Preparation of stromal cells highly expressing SCR-3
  AGM-s3-A7 cells in which SCR-3 was highly expressed
  were prepared by using the above retrovirus vector in
  the same manner as (2) of Example 2.
- 15 (3) Determination of activity to support hematopoietic stem cells of stromal cells in which SCR-3 is highly expressed

In the same manner as described in (III) (2) of Example 1, determination of the activity to support

hematopoietic stem cells was performed except that AGM-S3-A7 cells, AGM-S3-A7 cells in which SCR-3 was highly expressed through retrovirus, and AGM-S3-A7 cells into which a control vector was introduced were seeded in a 24-well culture dish (Falcon) at 1 × 10<sup>5</sup> cells/well.

25 The results are shown in Fig. 6. Hematopoietic cells co-cultured with AGM-s3-A7 cells in which SCR-3 was highly expressed (A7/SCR-3) showed high chimerism in

recipient individuals after the transplantation compared with the parent cell lines or hematopoietic cells cocultured with the cells into which a control vector was introduced. The high chimerism was observed in myeloid 5 and lymphoid cells two months after the transplantation. Therefore, it is revealed that hematopoietic stem cells and hematopoietic progenitor cells which can reconstitute the hematopoietic system in bodies of irradiated mice have maintained and amplified superiorly 10 to the co-culture with cells into which SCR-3 is not introduced, during the co-culture period. From the results, it is revealed that an activity of stromal cells to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor 15 cells is increased by high expression of SCR-3. Therefore, it is revealed that a gene product of SCR-3 has an activity to affect hematopoietic stem cells or hematopoietic progenitor cells to support survival or proliferation thereof or an activity to affect stromal 20 cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

# Example 4 Cloning of SCR-4 and activity determination

By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 3 with BLAST, it has been found that SCR-4 has a high homology to Homo sapiens

clone 25077 mRNA of an accession number AF131820, and that SCR-4 is a mouse ortholog. This sequence is described in WO 00/66784.

The nuclotide sequence of ORF of AF131820 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 16. Only the amino acid sequence is shown in SEQ ID NO: 17.

The nuclotide sequence of ORF of SCR-4 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 14. Only the amino acid sequence is shown in SEQ ID NO: 15.

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Determination of the activity of SCR-4 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

15 (1) Construction of retrovirus vector for expression of human SCR-4

From 3  $\mu g$  of mRNA derived from fetal liver (CLONETEC, U.S.A.), cDNA was synthesized by using oligo-dT primer and reverse transcriptase (SuperscriptII, GIBCO-BRL).

- Using the cDNA as a template, the ORF region of human SCR-4 was amplified by PCR with HSCR-4FxhoI and HSCR-4RecoRV primers having the following nucleotide sequences. An amplified fragment was digested with XhoI and inserted to the retrovirus vector pMX-IRES-GFP in
- 25 the same manner as described in (1) of Example 2. For the insertion, the pMX-IRES-GFP was digested with a restriction enzyme *EcoRI*, blunt-ended with KOD DNA

synthase (TOYOBO, Japan) and digested with a restriction enzyme XhoI.

HSCR-4FxhoI

CCGCTCGAGCCACCatgttggctgcaaggctggtgt (SEQ ID NO: 34)

5 HSCR-4RecoRV

CCGGATATCtcatttctttctqttqcctcca (SEQ ID NO: 35)

- (2) Preparation of stromal cells highly expressing human SCR-4
- AGM-s3-A9 cells in which human SCR-4 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
  - (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing human SCR-4, and
- 15 determination of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay

In the same manner as described in (III) (1) 3) to
4) of Example 1, AGM-s3-A9 cells in which SCR-4 was
20 highly expressed through retrovirus, AGM-s3-A9 cells
into which a control vector was introduced, or AGM-s3-A9
cells were co-cultured with CD34-positive hematopoietic
stem cells derived from human cord blood, and
proliferation statuses of hematopoietic stem cells and
25 hematopoietic progenitor cells are determined.

Fig. 6 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9

cells in which human SCR-4 was highly expressed, AGM-S3-A9 cells into which a control vector was introduced or AGM-S3-A9 cells for two weeks. As a result, the coculture with AGM-S3-A9 cells in which human SCR-4 was highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 increases by allowing human SCR-4 to be highly expressed. From the results, it has been revealed that human SCR-4 has 10 an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells or an activity to affect stromal cells to impart a hematopoietic cell-supporting activity to the stromal 15 cells.

### Example 5 Cloning of SCR-5 and activity determination

In the nucleotide sequence of SEQ ID NO: 4 obtained by the SBH analysis, the presence of ORF was predicted. The nucleotide sequence of ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 18. Only the amino acid sequence is shown in SEQ ID NO: 19.

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By searching GenBank database for the nucleotide

25 sequence of SEQ ID NO: 18 with BLAST, it has been found
that SCR-5 has a high homology with Homo sapiens
esophageal cancer related gene 4 portein (ECRG4) mRNA of

an accession number AF325503, and that SCR-5 is a mouse ortholog of AF325503. The nuclotide sequence of ORF of AF325503 and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 20. Only the amino acid sequence is shown in SEQ ID NO: 21.

Determination of the activity of SCR-5 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of nouse SCR-5

Based on the nucleotide sequence of SCR-5 ORF, SCR-5FxhoI and SCR-5Rblunt primers having the following nucleotide sequences were prepared for retrovirus cloning, and PCR was performed using DNA having the nucleotide sequence shown in SEQ ID NO: 23 as a template. An amplified fragment was digested with a restriction enzyme XhoI and inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in (1) of Example 2. For the insertion, the pMX-IRES-GFP was digested with a restriction enzyme EcoRI, blunt-ended with KOD DNA synthase (TOYOBO, Japan) and digested with a restriction enzyme XhoI.

SCR-5FxhoI

ccgCTCGAGccaccatgagcacctcgtctqcqcq (SEQ ID NO: 36)

25 SCR-5Rblunt

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tccGTTAACttaatagtcatcatagttca (SEQ ID NO: 37)

- (2) Preparation of stromal cells highly expressing SCR-5 AGM-s3-A7 cells in which SCR-5 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
- 5 (3) Determination of activity to support hematopoietic stem cells of stromal cells in which SCR-5 is highly expressed

In the same manner as described in (3) of Example 3, determination of the activity to support

10 hematopoietic stem cells was performed.

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The results are shown in Fig. 8. Hematopoietic cells co-cultured with AGM-s3-A7 cells in which SCR-5 was highly expressed (A7/SCR-5) showed high chimerism in recipient individuals after the transplantation compared with the parent cell lines or hematopoietic cells cocultured with the cells into which a control vector was introduced. The high chimerism was observed in myeloid and lymphoid cells two months after the transplantation. Therefore, it is revealed that hematopoietic stem cells and hematopoietic progenitor cells which can reconstitute the hematopoietic system in bodies of irradiated mice have maintained and amplified superiorly to the co-culture with cells into which SCR-5 is not introduced, during the co-culture period. From the results, it is revealed that an activity of stromal cells to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor

cells is increased by high expression of SCR-5.

Therefore, it is revealed that a gene product of SCR-5 has an activity to affect hematopoietic stem cells or hematopoietic progenitor cells to support survival or proliferation thereof or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

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## 10 Example 6 Cloning of SCR-6 and activity determination

Based on the nucleotide sequence of SEQ ID NO: 5, a probe was prepared and AGM-s3-A9 cDNA was screened by hybridization to obtain a gene containing ORF of mouse SCR-6.

AGM-s3-A9 cells (1.4 x 108 cells) were dissolved in 15 20 mL of ISOGEN (Nippon gene, Japan) and total RNAs were prepared according to the attachment. Messenger RNAs were prepared from one milligram of the total RNAs according to the protocol of the mRNA purification kit 20 (Amersham Pharmacia, U.S.A.). By using SMART cDNA library construction kit (CLONTECH, U.S.A.), cDNA libraries devided to 15 fractions were prepared from the 2 mg of the prepared mRNAs according to the attachment. The libraries contained about 400,000 of independent clones in total. For each fraction, PCR was performed 25 under the following conditions to identify a fraction containing SCR-6 cDNA.

Based on the sequence of a partial fragment of the mouse SCR-6 gene, the following primers were prepared, and PCR was performed with 35 cycles of 94°C, 30 seconds, 55°C, 30 seconds and 72°C, 1 minute, by using each

5 fraction of AGM-s3-A9 cDNA libraries as a template. SCR-6F

AGCTCATTACTGTATATTA (SEQ ID NO: 22; 1983-2002)
(SEQ ID NO: 38)
SCR-6R

10 GCTATATTTCATAAGTCATC (SEQ ID NO: 22; 2342-2361)
(SEQ ID NO: 39)

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The PCR product was subjected to 2% agarose gel electrophoresis and a fraction from which the PCR product having the expected size was obtained was identified. For each of two fractions among the positive fractions, 50,000 plaques were seeded on two 15-cm petri dishes and incubated 37°C for 10 hours. Then, plaques of each petri dish were replicated to a sheet of Biodyne nylon filter (Pall, U.S.A.). The replicated nylon filter was subjected to DNA fixation treatment according to the attachment, and screening with <sup>32</sup>P-labeled DNA probe was performed.

The probe was prepared as follows. PCR was

25 performed with 35 cycles of of 94°C, 30 seconds, 55°C,

30 seconds and 72°C, 1 minute, by using SCR-6F and SCR6R and the plasmid containing a partial fragment of the

mouse SCR-6 gene as a template. The PCR product was subjected to 2% agarose gel electrophoresis and the amplified fragment was purified by JETSORB. By using 25 ng of the obtained PCR fragment, <sup>32</sup>P-labeled DNA probe was prepared with Megaprime labeling kit (Amersham Pharmacia, U.S.A.).

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Hybridization and washing were performed with ExpressHybSolution (CLONETECH, U.S.A.) according to the attachment. An X-ray film was exposed to the filter and 10 developed with a Fuji film auto developer to analyze the result. A plaque at a position corresponding to the resultant strongly exposed portion was scraped from the petri dish, and seeded again so that about 200 of plaques should appear on 10-cm petri dish. Screening was again performed according to the above-mentioned 15 method to isolate a single plaque. The obtained clone was transfected to E. coli strain BM25.8 according to the attachment of SMART cDNA library construction kit, and the strain BM25.8 was cultured on in vivo LB agar medium to form colony. A single colony of the 20 transfected E. coli was inoculated to 3 ml of LB medium containing 50 µg/ml ampicilin and cultured at 30°C overnight. Plasmid was extracted with RPM kit (BIO101, U.S.A.) to obtain about 10 mg of plasmid.

25 Sequencing the both ends of the inserted fragment with an ABI377 DNA sequencer by using λTriplEx5'LDInsert Screening Amplimer (CTCGGGAAGCGCGCCATTGTGTTGGT

(SEQ ID NO: 40); CLONTECH, U.S.A.) revealed that it included cDNA containing the nucleotide sequence from nucleotide 1 of SEQ ID NO: 5. The full-length nucleotide sequence was also determined with the ABI377

- 5 DNA sequencer. The nuclotide sequence and the amino acid sequence deduced from a nucleotide sequence predicted as ORF in the nucleotide sequence are shown in SEQ ID NO: 22. Only the amino acid sequence is shown in SEQ ID NO: 23.
- Determination of the activity of SCR-6 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.
  - (1) Construction of retrovirus vector for expression of mouse SCR-6
- Based on the nucleotide sequence of SCR-6 ORF, SCR-6FxhoI and SCR-6Reco primers having the following sequences were prepared for retrovirus cloning, and PCR was performed by using DNA having the nucleotide sequence shown in SEQ ID NO: 22 as a template. An amplified fragment was inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in (1) of Example 2.

SCR-6FxhoI

ccqctcqaqccaccATGCGTTTTTGCCTCTTCTC (SEQ ID NO: 41)

25 SCR-6Reco

cqqaattcTTATTGGTTCACTCTGTCTG (SEQ ID NO: 42)

- (2) Preparation of stromal cells highly expressing SCR-6 AGM-s3-A9 cells in which SCR-6 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
- 5 (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-6, and determination of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay

In the same manner as described in (III) (1) 3) to

4) of Example 1, AGM-s3-A9 cells in which SCR-6 was
highly expressed through retrovirus, AGM-s3-A9 cells
into which a control vector was introduced, or AGM-s3-A9
cells were co-cultured with CD34-positive hematopoietic
stem cells derived from human cord blood, and

proliferation statuses of hematopoietic stem cells and
hematopoietic progenitor cells are determined.

Fig. 9 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9 cells in which SCR-6 was highly expressed (A9/SCR-9),

20 AGM-S3-A9 cells into which a control vector was introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two weeks. As a result, the co-culture with AGM-S3-A9 cells in which SCR-6 was highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been

25 revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 increases by allowing SCR-6 to be highly expressed.

From the results, it has been revealed that the gene product of SCR-6 has an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

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## Example 7 Cloning of SCR-7 and activity determination

In the nucleotide sequence of SEQ ID NO: 6 obtained by the SBH analysis, the presence of ORF was predicted.

The nuclotide sequence of ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 24. Only the amino acid sequence is shown in SEQ ID NO: 25.

Determination of the activity of SCR-7 to support the hematopoietic stem cells or hematopoietic progenitor cells was performed as follows.

(1) Construction of retrovirus vector for expression of 20 mouse SCR-7

Based on the nucleotide sequence of SCR-7 ORF, SCR-7Fsall and SCR-7Reco primers having the following nucleotide sequences were prepared for retrovirus cloning, and PCR was performed using DNA having the nucleotide sequence shown in SEQ ID NO: 24 as a template. An amplified fragment was inserted to the retrovirus vector pMX-IRES-GFP in the same manner as described in

- (1) of Example 2.
- SCR-7FSalI

10

acgcgtcgacccaccATGCCCCGCTACGAGTTG (SEQ ID NO: 43) SCR-7Reco

- 5 attGAATTCTCACTTCTTCCTCCTCTTTG (SEQ ID NO: 44)
  - (2) Preparation of stromal cells highly expressing SCR-7 AGM-s3-A9 cells in which SCR-7 was highly expressed were prepared by using the above retrovirus vector in the same manner as (2) of Example 2.
  - (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-7, and determination of proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells by clonogenic assay
- In the same manner as described in (III) (1) 3) to
  4) of Example 1, AGM-s3-A9 cells in which SCR-7 was
  highly expressed through retrovirus, AGM-s3-A9 cells
  into which a control vector was introduced, or AGM-s3-A9
  cells were co-cultured with CD34-positive hematopoietic
  stem cells derived from human cord blood, and
  proliferation statuses of hematopoietic stem cells and
  hematopoietic progenitor cells are determined.
- Fig. 10 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9 cells in which SCR-7 was highly expressed (A9/SCR-7), AGM-S3-A9 cells into which a control vector was introduced (A9/pMXIG) or AGM-S3-A9 cells (A9) for two

weeks. As a result, the co-culture with AGM-S3-A9 cells in which SCR-7 was highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 increases by allowing SCR-7 to be highly expressed. From the results, it has been revealed that the gene product of SCR-7 has an activity to support survival or proliferation of hematopoietic stem cells or lematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal cells or impart the activity to the stromal cells.

## 15 Example 8 Cloning of SCR-8 and activity determination

By searching GenBank database for the nucleotide sequence shown in SEQ ID NO: 7 with BLAST, it has been found that SCR-8 is the same gene as Mus musculus mRNA for ADAM23 of an accession number AB009673. The nuclotide sequence of SCR-8 ORF and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 26. Only the amino acid sequence is shown in SEQ ID NO: 27.

Also, the sequence encoding Human MDC3 protein [Homo sapiens] described by JP 11155574-A has a homology of not less than 90% with SCR-8 and, therefore, is a human ortholog of SCR-8. The nuclotide sequence of this ORF

and the amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 28. Only the amino acid sequence is shown in SEQ ID NO: 29.

Determination of the activity of SCR-8 to support

the hematopoietic stem cells or hematopoietic progenitor
cells was performed as follows.

(1) Construction of retrovirus vector for expression of mouse SCR-8

Based on the nucleotide sequence of SCR-8 ORF, SCR10 8FxhoI and SCR-8Reco primers having the following
nucleotide sequences were prepared, and PCR was
performed using AGM-s3-A9 cDNA as a template. An
amplified fragment was inserted to the retrovirus vector
pMX-IRES-GFP in the same manner as described in (1) of

15 Example 2.

SCR-8FxhoI

ccgctcgagccaccATGAAGCCGCCCGGCAGCATC (SEQ ID NO: 45)
SCR-8Reco

cggaattcTCAGATGGGGCCTTGCTGAGT (SEQ ID NO: 46)

20

- (2) Preparation of stromal cells highly expressing SCR-8
  AGM-s3-A9 cells in which SCR-8 was highly expressed
  were prepared by using the above retrovirus vector in
  the same manner as (2) of Example 2.
- 25 (3) Co-culture of human hematopoietic stem cells and stromal cells highly expressing SCR-8, and determination of proliferation statuses of hematopoietic stem cells

and hematopoietic progenitor cells by clonogenic assay

In the same manner as described in (III) (1) 3) to 4) of Example 1, AGM-s3-A9 cells in which SCR-8 was highly expressed through retrovirus, AGM-s3-A9 cells into which a control vector was introduced, or AGM-s3-A9 cells were co-cultured with CD34-positive hematopoietic stem cells derived from human cord blood, and proliferation statuses of hematopoietic stem cells and hematopoietic progenitor cells are determined.

3

10 Fig. 11 shows results when the CD34-positive hematopoietic stem cells were co-cultured with AGM-S3-A9 cells in which SCR-8 was highly expressed, AGM-S3-A9 cells into which a control vector was introduced or AGM-S3-A9 cells for two weeks. As a result, the co-15 culture with AGM-S3-A9 cells in which SCR-8 was highly expressed, increases of BFU-E and CFU-C were observed. Therefore, it has been revealed that the activity to support hematopoietic stem cells or hematopoietic progenitor cells, of AGM-S3-A9 increases by allowing 20 SCR-8 to be highly expressed. From the results, it has been revealed that the gene product of SCR-8 has an activity to support survival or proliferation of hematopoietic stem cells or hematopoietic progenitor cells or an activity to affect stromal cells to enhance a hematopoietic cell-supporting activity of the stromal 25 cells or impart the activity to the stromal cells.

## SEQUENCE LISTING

## <110> KIRIN BEER KABUSHIKI KAISHA

<12O> POLYPEPTIDE HAVING AN ACTIVITY TO SUPPORT PROLIFERATION OR SURVIVAL OF HEMATOPOIETIC STEM CELL AND HEMATOPOIETIC PROGENITOR CELL, AND DNA CODING FOR THE SAME

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cag Gln	ctg Leu	cac His	ccc Pro 180	cag Gln	ctg Leu	ctg Leu	ctg Leu	cct Pro 185	gat Asp	gac Asp	tac Tyr	ctg Leu	gac Asp 190	tgc Cys	ctg Leu	576
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					acc Thr											672
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					gtc Val											816
					ggc Gly											864
					ctg Leu											912
					gtg Val 310											960
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					acg Thr											1152
					cag Gln 390											1200
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ctg Leu	gcc Ala	aac Asn 435	cag Gln	atc Ile	aac Asn	aac Asn	ccc Pro 440	gag Glu	gtg Val	gag Glu	gtg Val	gac Asp 445	atc Ile	acc Thr	aag Lys	1344
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					ctc Leu											1584
					tgc Cys											1632
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Gly Tyr Thr Cys Cys Thr Ser Glu Met Glu Glu Asn Leu Ala Asn Arg 65 70 75 80

Ser His Ala Glu Leu Glu Thr Ala Leu Arg Asp Ser Ser Arg Val Leu 85 90 95

Gln Ala Met Leu Ala Thr Gln Leu Arg Ser Phe Asp Asp His Phe Gln 100 105 110

His Leu Leu Asn Asp Ser Glu Arg Thr Leu Gln Ala Thr Phe Pro Gly 115 120 125

Ala Phe Gly Glu Leu Tyr Thr Gln Asn Ala Arg Ala Phe Arg Asp Leu 130 135 140

Tyr Ser Glu Leu Arg Leu Tyr Tyr Arg Gly Ala Asn Leu His Leu Glu 145 150 155 160

Glu Thr Leu Ala Glu Phe Trp Ala Arg Leu Leu Glu Arg Leu Phe Lys 165 170 175

Gîn Leu His Pro Gîn Leu Leu Leu Pro Asp Asp Tyr Leu Asp Cys Leu 180 185 190

- Gly Lys Gln Ala Glu Ala Leu Arg Pro Phe Gly Glu Ala Pro Arg Glu 195 200 205
- Leu Arg Leu Arg Ala Thr Arg Ala Phe Val Ala Ala Arg Ser Phe Val 210 215 220
- Gln Gly Leu Gly Val Ala Ser Asp Val Val Arg Lys Val Ala Gln Val 225 230 235 240
- Pro Leu Gly Pro Glu Cys Ser Arg Ala Val Met Lys Leu Val Tyr Cys 245 250 255
- Ala His Cys Leu Gly Val Pro Gly Ala Arg Pro Cys Pro Asp Tyr Cys 260 265 270
- Arg Asn Val Leu Lys Gly Cys Leu Ala Asn Gln Ala Asp Leu Asp Ala 275 280 285
- Glu Trp Arg Asn Leu Leu Asp Ser Met Val Leu Ile Thr Asp Lys Phe 290 295 300
- Trp Gly Thr Ser Gly Val Glu Ser Val Ile Gly Ser Val His Thr Trp 305 310 315 320
- Leu Ala Glu Ala Ile Asn Ala Leu Gln Asp Asn Arg Asp Thr Leu Thr 325 330 335
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- Pro Gly Pro Glu Glu Lys Arg Arg Gly Lys Leu Ala Pro Arg Glu 355 360 365
- Arg Pro Pro Ser Gly Thr Leu Glu Lys Leu Val Ser Glu Ala Lys Ala 370 380
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Asn	Gly	Met	Ala 420	Arg	Gly	Arg	Tyr	Leu 425	Pro	Glu	Val	Met	G1 y 430	Asp	Gly	
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Pro	Asp 450	Met	Thr	Ile	Arg	G1n 455	Gln	Ile	Met	Gln	Leu 460	Lys	Пе	Met	Thr	
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ctt Leu	gga Gly	atc He	tgg Trp 20	gcc Ala	cag Gln	atc Ile	aca Thr	cat His 25	gca Ala	aca Thr	gag Glu	aca Thr	aaa Lys 30	gaa Glu	gtc Val	96

Gln Ser Ser Leu Lys Ala Gln Gln Gly Leu Glu Ile Glu Met Phe His 35 40 45	144
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Gln Ser Ser Leu Lys Ala Gln Gln Gly Leu Glu Ile Glu Met Phe His 35  Met Gly Phe Gln Asp Ser Ser Asp Cys Cys Leu Ser Tyr Asn Ser Arg	
20 25 30  Gln Ser Ser Leu Lys Ala Gln Gln Gly Leu Glu Ile Glu Met Phe His 35  Met Gly Phe Gln Asp Ser Ser Asp Cys Cys Leu Ser Tyr Asn Ser Arg 50  Ile Gln Cys Ser Arg Phe Ile Gly Tyr Phe Pro Thr Ser Gly Gly Cys	

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aca c Thr L																161
cca c Pro L	:tt .eu	gtg Val	aag Lys 30	aat Asn	tcc Ser	atc Ile	aca Thr	aag Lys 35	aac Asn	caa G1n	tgg Trp	ctc Leu	gta Val 40	aca Thr	ccc Pro	209
agc a Ser A	irg															257
act g Thr G	iga ily iO	caa G1n	gaa Glu	ctg Leu	aaa Lys	gag Glu 65	gca Ala	gcc Ala	ttg Leu	gaa Glu	cca Pro 70	tca Ser	atg Met	gaa Glu	aaa Lys	305
atc t Ile P 75																353
gct g Ala V																401
gag a Glu I																449
gat a Asp A																497
aca g	ct	ttg	tct	gcc	ttg	gca	gta	gcc	aga	aca	cct	gct	ctc	atg	aac	545

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		gga Gly														641
cca Pro	ggc Gly	cca Pro	aag Lys 190	cat His	ctg Leu	gct Ala	tgg Trp	atg Met 195	ctg Leu	cat His	tct Ser	ggt Gly	gtg Val 200	atg Met	ggt Gly	689
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		ctg Leu														881
		acc Thr														929
		tta Leu 285											Thr			977
		aaa Lys														1025
		atc Ile														1073
ttt Phe	atg Met	cga Arg	gtt Val	gca Ala 335	act Thr	atg Met	cta Leu	gca Ala	act Thr 340	gga Gly	agc Ser	aac Asn	aga Arg	aag Lys 345	aaa Lys	1121
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Lys Thr Arg Ile Arg Thr His Arg Gly Lys Thr Gly Gln Glu Leu Lys 50 55 60

Glu Ala Ala Leu Glu Pro Ser Met Glu Lys Ile Phe Lys Ile Asp Gln 65 70 75 80

Met Gly Arg Trp Phe Val Ala Gly Gly Ala Ala Val Gly Leu Gly Ala 85 90 95

Leu Cys Tyr Tyr Gly Leu Gly Met Ser Asn Glu Ile Gly Ala Ile Glu 100 105 110

Lys Ala Val Ile Trp Pro Gln Tyr Val Lys Asp Arg Ile His Ser Thr 115 120 125

Tyr Met Tyr Leu Ala Gly Arg Tyr Cys Leu Thr Ala Leu Ser Ala Leu 130 135 140

Ala Val Ala Arg Thr Pro Ala Leu Met Asn Phe Met Met Thr Gly Ser 145 150 155 160

Trp Val Thr Ile Gly Ala Thr Phe Ala Ala Met Ile Gly Ala Gly Met 165 170 175

Leu Val His Ser Ile Ser Tyr Glu Gln Ser Pro Gly Pro Lys His Leu 180 185 190

Ala Trp Met Leu His Ser Gly Val Met Gly Ala Val Val Ala Pro Leu 195 200 205

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Gly 225	He	Val	Gly	Gly	Leu 230	Ser	Thr	Val	Ala	Met 235	Cys	Ala	Pro	Ser	Glu 240	
Lys	Phe	Leu	Asn	Met 245	Gly	Ala	Pro	Leu	Gly 250	Val	Gly	Leu	Gly	Leu 255	Val	
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													tec Ser			384
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													cat His 190			576
													cct Pro			624
ata Ile	tta Leu 210	999 Gly	ggt Gly	cct Pro	ctt Leu	ctc Leu 215	atc Ile	aga Arg	gct Ala	gca Ala	tgg Trp 220	tac Tyr	aca Thr	gct Ala	ggc Gly	672
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atg ttc ctt Met Phe Leu 290	ctg tat gai Leu Tyr Asi	acc cag aa Thr Gln Ly 295	ys Val Ile	aag cgt gca Lys Arg Ala 300	gaa gta 912 Glu Val
tca cca atg Ser Pro Met 305	tat gga gti Tyr Gly Va 310	Gln Lys Ty	at gat ccc yr Asp Pro 315	att aac tcg Ile Asn Ser	atg ctg 960 Met Leu 320
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Cys Tyr Tyr Gly Leu Gly Leu Ser Asn Glu Ile Gly Ala Ile Glu Lys

100 105 110

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Met Tyr Leu Ala Gly Ser Ile Gly Leu Thr Ala Leu Ser Ala Ile Ala 130 135 140

Ile Ser Arg Thr Pro Val Leu Met Asn Phe Met Met Arg Gly Ser Trp 145 150 155 160

Val Thr Ile Gly Val Thr Phe Ala Ala Met Val Gly Ala Gly Met Leu 165 170 175

Val Arg Ser Ile Pro Tyr Asp Gln Ser Pro Gly Pro Lys His Leu Ala 180 185 190

Trp Leu Leu His Ser Gly Val Met Gly Ala Val Val Ala Pro Leu Thr 195 200 205

Ile Leu Gly Gly Pro Leu Leu Ile Arg Ala Ala Trp Tyr Thr Ala Gly 210 215 220

Ile Val Gly Gly Leu Ser Thr Val Ala Met Cys Ala Pro Ser Glu Lys 225 230 240

Phe Leu Asn Met Gly Ala Pro Leu Gly Val Gly Leu Gly Leu Val Phe 245 250 255

Val Ser Ser Leu Gly Ser Met Phe Leu Pro Pro Thr Thr Val Ala Gly 260 265 270

Ala Thr Leu Tyr Ser Val Ala Met Tyr Gly Gly Leu Val Leu Phe Ser 275 280 285

Met Phe Leu Leu Tyr Asp Thr Gln Lys Val Ile Lys Arg Ala Glu Val 290 295 300

Ser Pro Met Tyr Gly Val Gln Lys Tyr Asp Pro Ile Asn Ser Met Leu 305 310 315 320

Ser Ile Tyr Met Asp Thr Leu Asn Ile Phe Met Arg Val Ala Thr Met

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Thr Asn Val Ala Val Ala Glu Asn Thr Ala Lys Glu Phe Leu Gly Gly 50 60

Leu Lys Arg Ala Lys Arg Gln Leu Trp Asp Arg Thr Arg Pro Glu Val 65 70 75 80

Gln Gln Trp Tyr Gln Gln Phe Leu Tyr Met Gly Phe Asp Glu Ala Lys 85 90 95

Phe Glu Asp Asp Val Asn Tyr Trp Leu Asn Arg Asn Arg Asn Gly His 100 105 110

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aaa Lys																144
act Thr																192
ctg Leu 65																240
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gaa Glu																384
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Phe Glu Asp Asp Ile Thr Tyr Trp Leu Asn Arg Asp Arg Asn Gly His 100 105 110

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gat tot tgc ttt agc aaa gac ttt tgt acg aag tgc aaa gta ggc ttt Asp Ser Cys Phe Ser Lys Asp Phe Cys Thr Lys Cys Lys Val Gly Phe 105 110 115	992
tat ttg cat aga ggc cgc tgc ttt gat gaa tgt cca gat ggt ttt gca Tyr Leu His Arg Gly Arg Cys Phe Asp Glu Cys Pro Asp Gly Phe Ala 120 125 130	1040
ccg tta gat gag act atg gaa tgt gta gaa ggt tgt gaa gtt ggt cat Pro Leu Asp Glu Thr Met Glu Cys Val Glu Gly Cys Glu Val Gly His 135 140 145	1088
tgg agc gaa tgg gga acg tgt agc aga aac aac cgc acg tgt gga ttt Trp Ser Glu Trp Gly Thr Cys Ser Arg Asn Asn Arg Thr Cys Gly Phe 150 155 160 165	1136
aaa tgg ggt ctg gaa acc aga aca cgg cag att gtt aaa aag cca gca Lys Trp Gly Leu Glu Thr Arg Thr Arg Gln Ile Val Lys Lys Pro Ala 170 175 180	1184
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Lys Asp Thr Ile Pro Cys Pro Thr Ile Ala Glu Ser Arg Arg Cys Lys 185 190 195	
atg gcc atg agg cac tgt cca gga gga aag aga aca cca aag gca aaa Met Ala Met Arg His Cys Pro Gly Gly Lys Arg Thr Pro Lys Ala Lys 200 205 210	1280
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gag cag cac agc gtc ttc ctc gct aca gac aga gtg aac caa Glu Gln His Ser Val Phe Leu Ala Thr Asp Arg Val Asn Gln 230 235 240	1370
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Asp Asn Gly Cys Ser Arg Cys Gln Gln Lys Leu Phe Phe Phe Leu Arg 50 60

Arg Glu Gly Met Arg Gln Tyr Gly Glu Cys Leu His Ser Cys Pro Ser 65 70 75 80

Gly Tyr Tyr Gly His Arg Ala Pro Asp Met Asn Arg Cys Ala Arg Cys 85 90 95

Arg Ile Glu Asn Cys Asp Ser Cys Phe Ser Lys Asp Phe Cys Thr Lys 100 105 110

Cys Lys Val Gly Phe Tyr Leu His Arg Gly Arg Cys Phe Asp Glu Cys 115 120 125

Pro Asp Gly Phe Ala Pro Leu Asp Glu Thr Met Glu Cys Val Glu Gly 130 135 140	
Cys Glu Val Gly His Trp Ser Glu Trp Gly Thr Cys Ser Arg Asn Asn 145 150 156 160	
Arg Thr Cys Gly Phe Lys Trp Gly Leu Glu Thr Arg Thr Arg Gln Ile 165 170 175	
Val Lys Lys Pro Ala Lys Asp Thr Ile Pro Cys Pro Thr Ile Ala Glu 180 185 190	
Ser Arg Arg Cys Lys Met Ala Met Arg His Cys Pro Gly Gly Lys Arg 195 200 205	
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caacctgggc g atg ccc cgc tac gag ttg gct ttg att ctg aaa gcc atg Met Pro Arg Tyr Glu Leu Ala Leu Ile Leu Lys Ala Met 1 5 10	170
cgg cgg cca gag acc gct gct ttg aaa cgt aca ata gaa tcc ctg Arg Arg Pro Glu Thr Ala Ala Ala Leu Lys Arg Thr Ile Glu Ser Leu 15 20 25	218

atg gac cga gga gcc ata gtg agg aac ttg gaa agc ctg ggt gag cgt Met Asp Arg Gly Ala Ile Val Arg Asn Leu Glu Ser Leu Gly Glu Arg 30 35 40 45	266
gcg ctc ccc tac agg atc tcg agt cac agc cag cag cac agc cga gga Ala Leu Pro Tyr Arg Ile Ser Ser His Ser Gln Gln His Ser Arg Gly 50 55 60	314
ggg tat ttc ctg gtg gat ttt tat gct ccg aca agt gct gtg gag aac Gly Tyr Phe Leu Val Asp Phe Tyr Ala Pro Thr Ser Ala Val Glu Asn 65 70 75	362
ata ctg gaa cac ttg gcg cga gac att gac gtg gtt aga cca aat att Ile Leu Glu His Leu Ala Arg Asp Ile Asp Val Val Arg Pro Asn Ile 80 85 90	410
gtg aaa cac cct ctg acc cag gaa gta aaa gag tgt gac ggc ata gtc Val Lys His Pro Leu Thr Gln Glu Val Lys Glu Cys Asp Gly Ile Val 95 100 105	458
cca gtc cca ctt gaa gaa aaa ctg tat tca aca aag agg agg aag aag Pro Val Pro Leu Glu Glu Lys Leu Tyr Ser Thr Lys Arg Arg Lys Lys 110 115 120 125	506
tgagaagatt caccagattc tggccttata tttaatccta agggcactat gggtgctgct	566
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Gly Ala Ile Val Arg Asn Leu Glu Ser Leu Gly Glu Arg Ala Leu Pro 35 40 45

Tyr Arg Ile Ser Ser His Ser Gln Gln His Ser Arg Gly Gly Tyr Phe

Leu 65	Val	Asp	Phe	Tyr	Ala 70	Pro	Thr	Ser	Ala	Va1 75	Glu	Asn	Ile	Leu	G1 u 80	
His	Leu	Ala	Arg	Asp 85	He	Asp	Val	Val	Arg 90	Pro	Asn	Ile	Val	Lys 95	His	
Pro	Leu	Thr	G1n 100	Glu	Val	Lys	Glu	Cys 105	Asp	Gly	Пе	Val	Pro 110	Val	Pro	
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agc Ser	ctt Leu	ccc Pro	ggc Gly 20	gcc Ala	tcc Ser	tgc Cys	ggc Gly	ccc Pro 25	ggc Gly	cgc Arg	tgc Cys	ccc Pro	gcc Ala 30	ggc ggc	ccg Pro	96
gtg Vai	ccg Pro	gcc Ala 35	cgc Arg	gcg Ala	ccg Pro	ccc Pro	tgc Cys 40	cgc Arg	ctg Leu	ctc Leu	ctc Leu	gtc Val 45	ctt Leu	ctc Leu	ctg Leu	144
cta Leu	cct Pro 50	gcg Ala	ctc Leu	gcc Ala	acc Thr	tca Ser 55	tcc Ser	cgg Arg	ccc Pro	cgt Arg	gcc Ala 60	cgg Arg	999 Gly	gcc Ala	gct Ala	192
		agc Ser														240
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		agc Ser														336

		gtg Val 115														384
		aca Thr														432
		cag Gln														480
ctt Leu	gac Asp	ctc Leu	aca Thr	ctg Leu 165	aac Asn	aat Asn	ggt Gly	ttg Leu	cta Leu 170	tct Ser	tct Ser	gac Asp	tac Tyr	gtg Val 175	gag Glu	528
		tat Tyr														576
tgt Cys	tac Tyr	tac Tyr 195	cac His	gga Gly	agc Ser	atc Ile	aga Arg 200	ggc Gly	gtc Val	aag Lys	gat Asp	tcc Ser 205	agg Arg	gtg Val	gct Ala	624
		acc Thr														672
gtg Val 225	tat Tyr	atg Met	ata Ile	gag Glu	cct Pro 230	ctg Leu	gaa Glu	ctg Leu	act Thr	gat Asp 235	gat Asp	gag Glu	aaa Lys	agc Ser	aca Thr 240	720
		ccg Pro														768
		aag Lys		Leu			Asp									816
		tta Leu 275														864
		gtg Val														912
		aag Lys														960
		gca Ala														1008

cag Gln	ctc Leu	aac Asn	acc Thr 340	agg Arg	gtt Val	gtc Val	Leu	gtg Val 345	gct Ala	gtc Val	gag Glu	acc Thr	tgg Trp 350	acc Thr	gag Glu	1	056
	gat Asp															1	104
Phe	tec Ser 370	aag Lys	tac Tyr	cgg Årg	cag G1n	cga Arg 375	atc Ile	aaa Lys	cag G1n	cac His	gct Ala 380	gac Asp	gcg Ala	gtc Val	cac His	1	152
	atc Ile															***************************************	200
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	ctt Leu															1	296
	ctt Leu															7	344
tgc Cys	ata Ile 450	gag Glu	tcc Ser	tgg Trp	ggc Gly	ggc Gly 455	tgc Cys	atc Ile	atg Met	gaa Glu	gaa Glu 460	aca Thr	999 G1 y	gtg Val	tcc Ser	1	392
	tct Ser															1	1440
	cag Gln				Gly			Leu								1	1488
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	gac Asp															1	1584
tgt Cys	tcg Ser 530	ctc Leu	tcc Ser	aat Asn	999 G1 y	gcc Ala 535	cac His	tgc Cys	agt Ser	gac Asp	ggc Gly 540	ccc Pro	tgc Cys	tgt Cys	aac Asn	1	1632
	acc Thr															1	1680

					atc Ile											1728
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					gaa Glu 630											1920
					ccg Pro											1968
					ctt Leu											2016
					act Thr											2064
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					ccg Pro 710											2160
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					tgc Cys											2256
					ttc Phe											2304
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cct agc gcc acc aat ctc ata ata ggc tcc atc gct ggt gcc atc ctg Pro Ser Ala Thr Asn Leu Ile Ile Gly Ser Ile Ala Gly Ala Ile Leu 785 790 795 800	2400
gta gca gct att gtc ctt ggg ggc aca ggc tgg gga ttt aaa aac gtc Val Ala Ala Ile Val Leu Gly Gly Thr Gly Trp Gly Phe Lys Asn Val 805 810 815	2448
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Leu Pro Ala Leu Ala Thr Ser Ser Arg Pro Arg Ala Arg Gly Ala Ala 50 55 60	
Ala Pro Ser Ala Pro His Trp Asn Glu Thr Ala Glu Lys Thr Leu Gly 65 70 75 80	
Val Leu Ala Asp Glu Asp Asn Thr Leu Gln Gln Asn Ser Ser Ser Arg 85 90 95	
Asn Thr Ser Tyr Ser Ser Ala Val Gln Lys Glu Ile Thr Leu Pro Ser 100 105 110	
Arg Leu Val Tyr Tyr Ile Asn Gln Asp Ser Glu Ser Pro Tyr His Val 115 120 125	
Leu Asp Thr Lys Ala Arg His Gln Gln Lys His Asn Lys Ala Val His 130 135 140	

Leu	Ala	Gln	Ala	Ser	Phe	Gln	Ile	Glu	Ala	Phe	Gly	Ser	Lys	Phe	He
145					150					155	_		-		160

Leu Asp Leu Thr Leu Asn Asn Gly Leu Leu Ser Ser Asp Tyr Val Glu 165 170 175

Ile His Tyr Glu Asp Gly Lys Gln Met Tyr Ser Lys Gly Gly Glu His 180 185 190

Cys Tyr Tyr His Gly Ser Ile Arg Gly Val Lys Asp Ser Arg Val Ala 195 200 205

Leu Ser Thr Cys Asn Gly Leu His Gly Met Phe Glu Asp Asp Thr Phe 210 220

Val Tyr Met Ile Glu Pro Leu Glu Leu Thr Asp Asp Glu Lys Ser Thr 225 230 235 240

Gly Arg Pro His Ile Ile Gln Lys Thr Leu Ala Gly Gln Tyr Ser Lys 245 250 255

Gin Met Lys Asn Leu Ser Thr Asp Gly Ser Asp Gln Trp Pro Leu Leu 260 265 270

Pro Glu Leu Gln Trp Leu Arg Arg Arg Lys Arg Ala Val Asn Pro Ser 275 280 285

Arg Gly Val Phe Glu Glu Met Lys Tyr Leu Glu Leu Met Ile Val Asn 290 295 300

Asp His Lys Thr Tyr Lys Lys His Arg Ser Ser His Ala His Thr Asn 305 310 315 320

Asn Phe Ala Lys Ser Val Val Asn Leu Val Asp Ser Ile Tyr Lys Glu 325 330 335

Gin Leu Asn Thr Arg Val Val Leu Val Ala Val Glu Thr Trp Thr Glu 340 345 350

Lys Asp His Ile Asp Ile Thr Ile Asp Pro Val Gln Met Leu His Asp 355 360 365

Phe Ser Lys Tyr Arg Gln Arg Ile Lys Gln His Ala Asp Ala Val His 370 380

Leu Ile Ser Arg Val Thr Phe His Tyr Lys Arg Ser Ser Leu Ser Tyr 385 390 395 400

Phe Gly Gly Val Cys Ser Arg Ile Arg Gly Val Gly Val Asn Glu Tyr 405 410 415

Gly Leu Pro Met Ala Val Ala Gln Val Leu Ser Gln Ser Leu Ala Gln 420 425 430

Asn Leu Gly Ile Gln Trp Glu Pro Ser Ser Arg Lys Pro Lys Cys Glu 435 440 445

Cys Ile Glu Ser Trp Gly Gly Cys Ile Met Glu Glu Thr Gly Val Ser 450 455 460

His Ser Arg Lys Phe Ser Lys Cys Ser Ile Leu Glu Tyr Arg Asp Phe 465 470 475 480

Leu Gln Arg Gly Gly Gly Ala Cys Leu Phe Asn Arg Pro Thr Lys Leu 485 490 495

Phe Glu Pro Thr Glu Cys Gly Asn Gly Tyr Val Glu Ala Gly Glu Glu 500 505 510

Cys Asp Cys Gly Phe His Val Glu Cys Tyr Gly Val Cys Cys Lys Lys 515 520 525

Cys Ser Leu Ser Asn Gly Ala His Cys Ser Asp Gly Pro Cys Cys Asn 530 540

Asn Thr Ser Cys Leu Phe Gln Ser Arg Gly Tyr Glu Cys Arg Asp Ala 545 550 555 560

Val Asn Ser Cys Asp Ile Thr Glu Tyr Cys Thr Gly Asp Ser Gly Gln 575

Cys Pro Pro Asn Leu His Lys Gln Asp Gly Tyr Ser Cys Asn Gln Asn 580 585 590

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- Gln Tyr Ile Trp Gly Thr Lys Ala Ala Gly Ser Asp Lys Phe Cys Tyr 610 615 620
- Glu Lys Leu Asn Thr Glu Gly Thr Glu Lys Gly Asn Cys Gly Lys Asp 625 630 635 640
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- Gly Glu Ile Ile Pro Thr Ser Phe Tyr His Gln Gly Arg Val Ile Asp 675 680 685
- Cys Ser Gly Ala His Val Val Leu Asp Asp Asp Thr Asp Val Gly Tyr 690 695 700
- Val Glu Asp Gly Thr Pro Cys Gly Pro Ser Met Met Cys Leu Asp Arg 705 710 715 720
- Lys Cys Leu Gln Ile Gln Ala Leu Asn Met Ser Ser Cys Pro Leu Asp 725 730 735
- Ser Arg Gly Lys Val Cys Ser Gly His Gly Val Cys Ser Asn Glu Ala 740 745 750
- Thr Cys Ile Cys Asp Phe Thr Trp Ala Gly Thr Asp Cys Ser Ile Arg 755 760 765
- Asp Pro Val Arg Asn Pro Asn Pro Pro Lys Asp Glu Gly Pro Lys Gly 770 780
- Pro Ser Ala Thr Asn Leu Ile Ile Gly Ser Ile Ala Gly Ala Ile Leu 785 790 795 800
- Val Ala Ala Ile Val Leu Gly Gly Thr Gly Trp Gly Phe Lys Asn Val 805 810 815

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gtc ctt ( Val Leu   50			Pro											70 (	92
tgg ggg g Trp Gly 7 65	get get Ala Ala	Alal	ccc Pro 70	agc Ser	gct Ala	ccg Pro	cat His	tgg Trp 75	aat Asn	gaa Glu	act Thr	gca Ala	gaa Glu 80	24	40
aaa aat Lys Asn I														28	88
agc agc a Ser Ser S		Ile :												30	36
ctg cct Leu Pro	tca aga Ser Arg 115	ctc a	ata Ile	tat Tyr	tac Tyr 120	atc Ile	aac Asn	caa Gln	gac Asp	tcg Ser 125	gaa Glu	agc Ser	cct Pro	38	84
tat cac Tyr His 130	gtt ctt Val Leu	gac a	aca Thr	aag Lys 135	gca Ala	aga Arg	cac His	cag G1n	caa G1n 140	aaa Lys	cat His	aat Asn	aag Lys	40	32
gct gtc Ala Val 145	cat ctg His Leu	Ala	cag G1n 150	gca Ala	agc Ser	ttc Phe	cag Gln	att 11e 155	gaa Glu	gcc Ala	ttc Phe	ggc Gly	tcc Ser 160	48	30
aaa ttc Lys Phe														52	28

165 170 175 tat gtg gag att cac tac gaa aat ggg aaa cca cag tac tct aag ggt 576 Tyr Val Glu Ile His Tyr Glu Asn Gly Lys Pro Gln Tyr Ser Lys Gly 185 gga gag cac tgt tac tac cat gga agc atc aga ggc gtc aaa gac tcc 624 Gly Glu His Cys Tyr Tyr His Gly Ser Ile Arg Gly Val Lys Asp Ser 200 aag gtg gct ctg tca acc tgc aat gga ctt cat ggc atg ttt gaa gat 672 Lys Val Ala Leu Ser Thr Cys Asn Gly Leu His Gly Met Phe Glu Asp 210 215 gat acc ttc gtg tat atg ata gag cca cta gag ctg gtt cat gat gag 720 Asp Thr Phe Val Tyr Met Ile Glu Pro Leu Glu Leu Val His Asp Glu 225 230 aaa agc aca ggt cga cca cat ata atc cag aaa acc ttg gca gga cag 768 Lys Ser Thr Gly Arg Pro His Ile Ile Gln Lys Thr Leu Ala Gly Gln 245 250 tat tot aag caa atg aag aat oto act atg gaa aga ggt gac cag tgg 816 Tyr Ser Lys Glin Met Lys Asn Leu Thr Met Gliu Arg Gly Asp Glin Trp ccc ttt ctc tct gaa tta cag tgg ttg aaa aga agg aag aga gca gtg 864 Pro Phe Leu Ser Glu Leu Gln Trp Leu Lys Arg Arg Lys Arg Ala Val 280 aat cca tca cgt ggt ata ttt gaa gaa atg aaa tat ttg gaa ctt atg 912 Asn Pro Ser Arg Gly Ile Phe Glu Glu Met Lys Tyr Leu Glu Leu Met 290 295 att gtt aat gat cac aaa acg tat aag aag cat cgc tot tot cat gca 960 Ile Val Asn Asp His Lys Thr Tyr Lys Lys His Arg Ser Ser His Ala 305 310 320 cat acc aac aac ttt gca aag tcc gtg gtc aac ctt gtg gat tct att 1008 His Thr Asn Asn Phe Ala Lys Ser Val Val Asn Leu Val Asp Ser Ile 325 330 335 tac aag gag cag ctc aac acc agg gtt gtc ctg gtg gct gta gag acc 1056 Tyr Lys Glu Gln Leu Asn Thr Arg Val Val Leu Val Ala Val Glu Thr 340 345 350 tgg act gag aag gat cag att gac atc acc acc aac cct gtg cag atg 1104 Trp Thr Glu Lys Asp Gln Ile Asp Ile Thr Thr Asn Pro Val Gln Met 360.ctc cat gag ttc tca aaa tac cgg cag cgc att aag cag cat gct gat 1152 Leu His Glu Phe Ser Lys Tyr Arg Gln Arg Ile Lys Gln His Ala Asp 370 375 get gtg cae etc ate teg egg gtg aca ttt cae tat aag aga age agt 1200 Ala Val His Leu Ile Ser Arg Val Thr Phe His Tyr Lys Arg Ser Ser

385	390	395	400
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	Arg Gly Gly Gly	gcc tgc ctt ttc aac Ala Cys Leu Phe Asn 490	
		gga aat gga tac gtg Gly Asn Gly Tyr Val 510	
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		get cac tgc agc gac Ala His Cys Ser Asp 540	
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	Glu Cys Asp Ile	act gaa tat tgt act Thr Glu Tyr Cys Thr 570	
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## WHAT IS CLAIMED IS:

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- 1. A DNA coding for a polypeptide of the following (A) or (B):
- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25; or
- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
  - 2. The DNA according to claim 1, which is a DNA of the following (a) or (b):
- 15 (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the nucleotide sequence of nucleotides 642 to 1370 of SEQ ID NO: 22, and the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24; or
  - (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
    - 3. The DNA according to claim 2, the stringent

condition is 6 x SSC, 5 x Denhardt, 0.5% SDS and 68°C (SSC: 3 M NaCl, 0.3 M sodium citrate; 50 x Denhardt: 1% BSA, 1% polyvinyl pyrrolidone, 1% Ficoll 400), or 6 x SSC, 5 x Denhardt, 0.5% SDS, 50% formamide and 42°C.

- 5 4. A expression vector which comprises the DNA of any one of claims 1 to 3 in such a manner that the DNA can be expressed.
- 5. A cell into which the DNA of any one of claims 1 to 3 is introduced in such a manner that the 10 DNA can be expressed.

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- 6. A polypeptide which is an expression product of the DNA of any one of claims 1 to 3, the polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- 7. The polypeptide according to claim 6, which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 23 and SEQ ID NO: 25, or an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence.
- 8. The polypeptide according to claim 6 or 7, which is modified with one or more modifying agents selected from the group consisting of polyethylene glycol (PEG), dextran, poly(N-vinyl-pyrrolidone), polypropylene glycol homopoymer, copolymer of polypropylene oxide/ethylene oxide, polyoxyethylated

polyol and polyvinyl alcohol.

- 9. An monoclonal antibody which binds to the polypeptide of any one of claims 6 to 8.
- 10. A method for supporting proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells, comprising the step of co-culturing stromal cells in which a DNA coding for a polypeptide of the following (A) or (B) is expressed, with hematopoietic stem cells or progenitor cells,
- (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
- 15 (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
  - 11. The method according to claim 10, wherein the DNA is a DNA of the following (a) or (b):
  - (a) a DNA which comprises a nucleotide sequence selected from the group consisting of the nucleotide sequence of nucleotides 1 to 1671 of SEQ ID NO: 8, the nucleotide sequence of nucleotides 1 to 1674 of SEQ ID NO: 10, the nucleotide sequence of nucleotides 1 to 366

of SEQ ID NO: 12, the nucleotide sequence of nucleotides 84 to 1121 of SEQ ID NO: 14, the nucleotide sequence of nucleotides 1 to 1035 of SEQ ID NO: 16, the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 18, the nucleotide sequence of nucleotides 1 to 444 of SEQ ID NO: 20, the nucleotide sequence of nucleotides 642 to 1370 of SEQ ID NO: 22, the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24, the nucleotide sequence of nucleotides 132 to 506 of SEQ ID NO: 24, the nucleotide sequence of nucleotides 1 to 2487 of SEQ ID NO: 26, and the nucleotide sequence of nucleotides 1 to 2496 of SEQ ID NO: 28; or

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- (b) a DNA which is hybridizable with a DNA comprising the nucleotide sequence as defined in (a) or a prove prepared from said DNA, under the stringent condition, and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- 12. A method for supporting proliferation or survival of hematopoietic stem cells or hematopoietic 20 progenitor cells, comprising the step of culturing hematopoietic stem cells or progenitor cells in the presence of a polypeptide of the following (A) or (B), said polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells when the hematopoietic stem cells or hematopoietic progenitor cells are cultured in the presence of the polypeptide,

(A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or

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- (B) a polypeptide which comprises an amino acid sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.
- effect to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor

  15 cells, which comprises an effective amount of a polypeptide of the following (A) or (B), said polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells when hematopoietic stem cells or hematopoietic or hematopoietic progenitor cells are cultured in the presence of the polypeptide,
  - (A) a polypeptide which comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, and SEQ ID NO: 29; or
    - (B) a polypeptide which comprises an amino acid

sequence including deletion, substitution or insertion of one or several amino acids in the amino acid sequence as defined in (A), and which has an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells.

## ABSTRACT OF THE DISCLOSURE

A gene encoding a polypeptide having an activity to support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is isolated by comparing expressed genes between cells which support proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells and cells which do not support the proliferation or survival. Proliferation or survival of hematopoietic stem cells or hematopoietic progenitor cells is supported by using stromal cells in which the isolated gene is expressed or a gene product of the isolated gene.

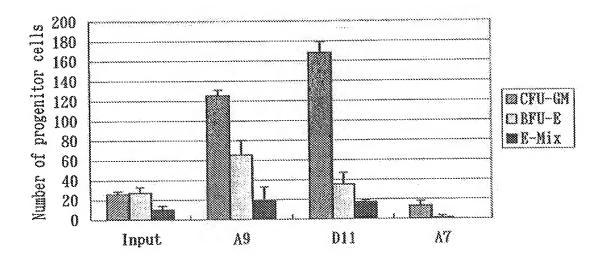


Fig.1

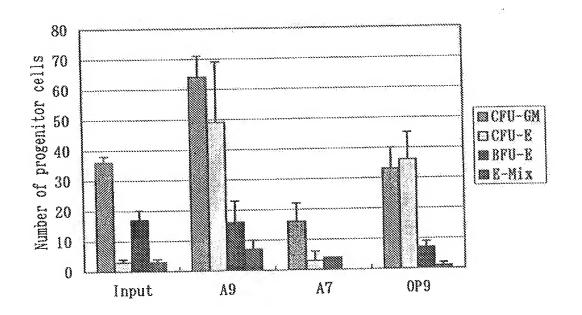
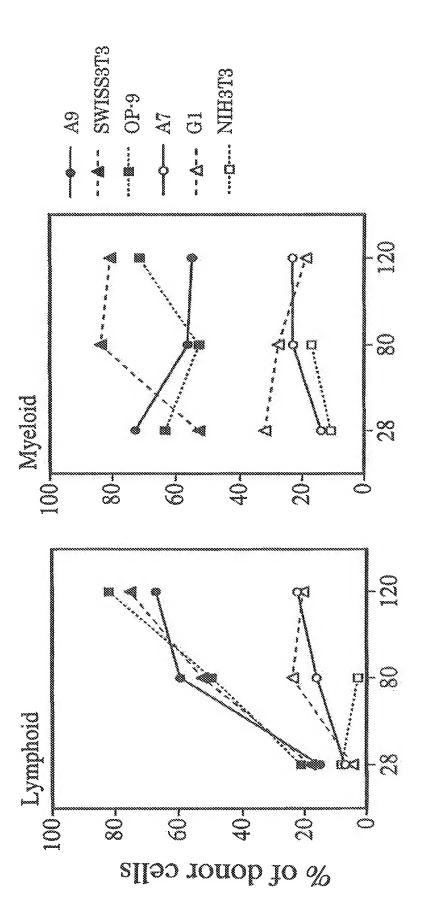
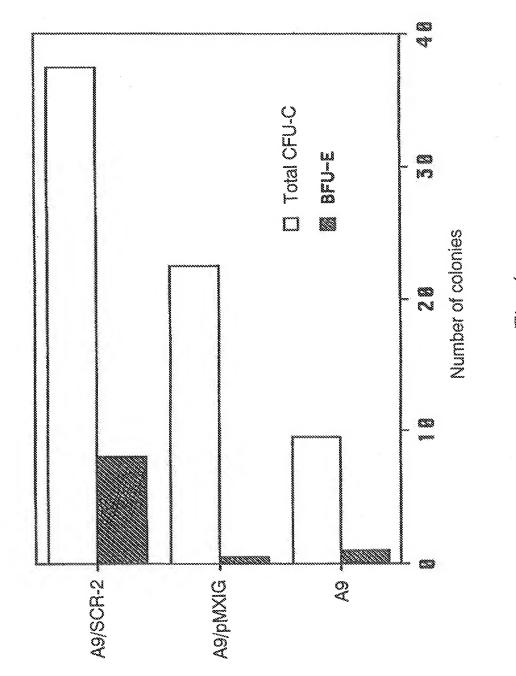


Fig.2

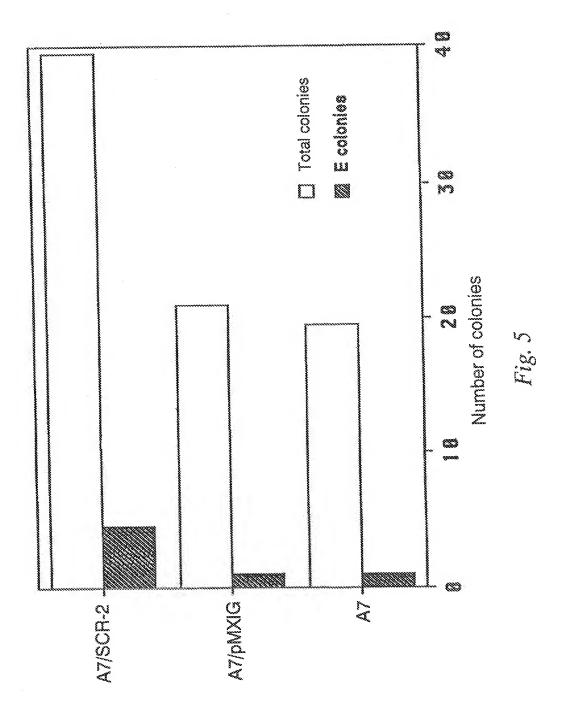


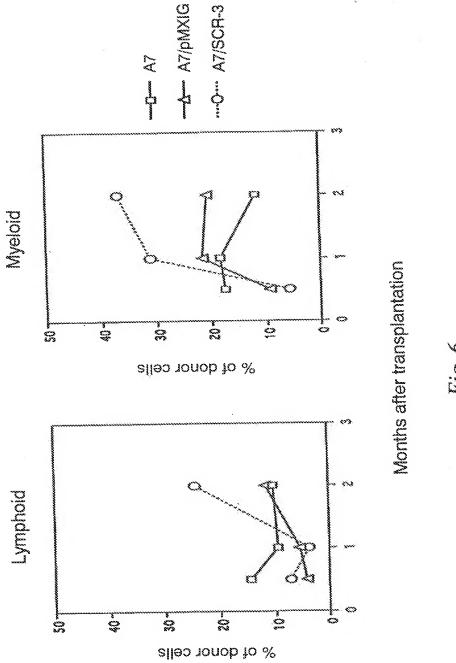
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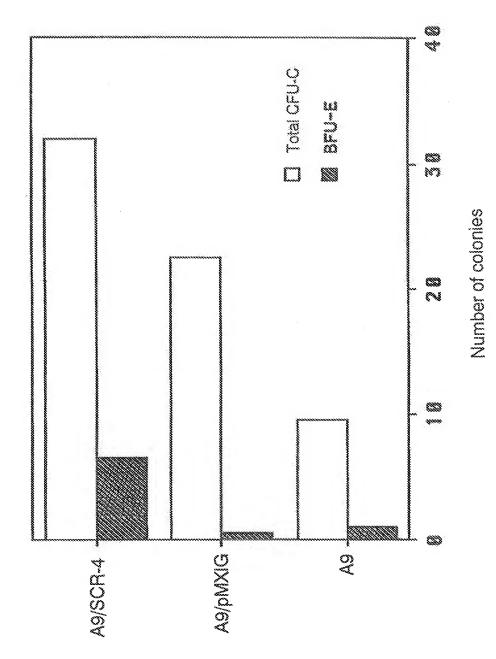
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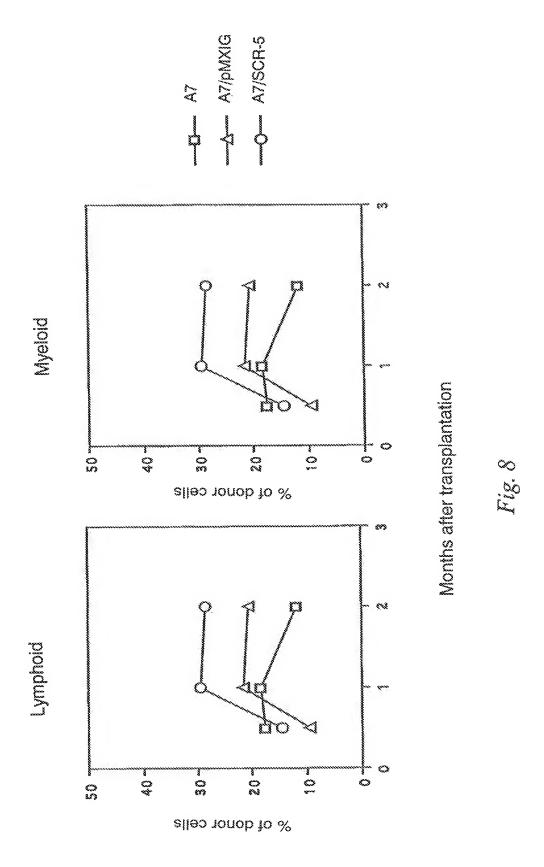
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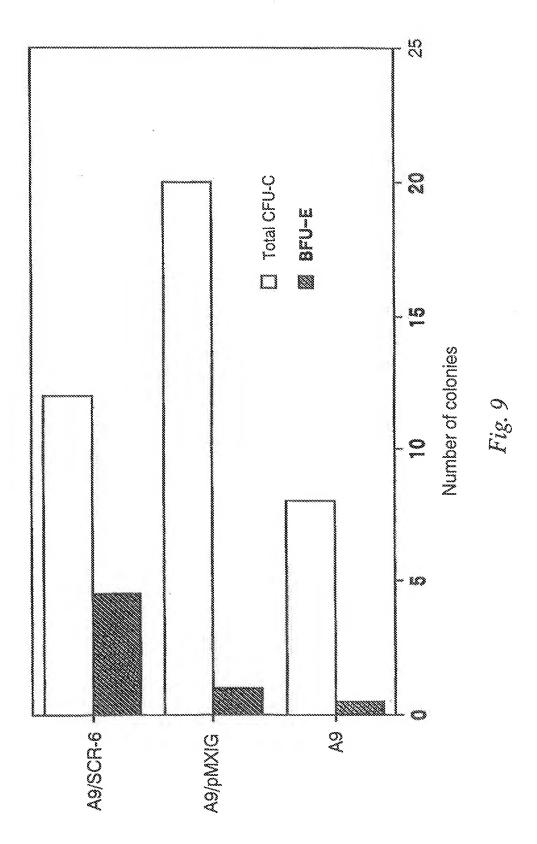


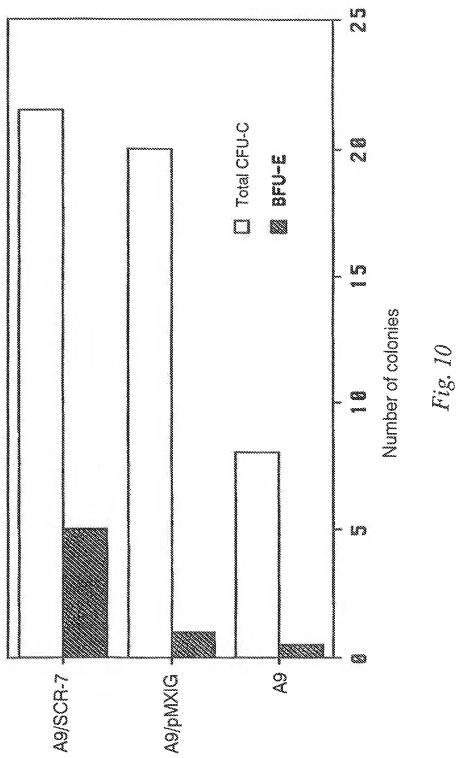


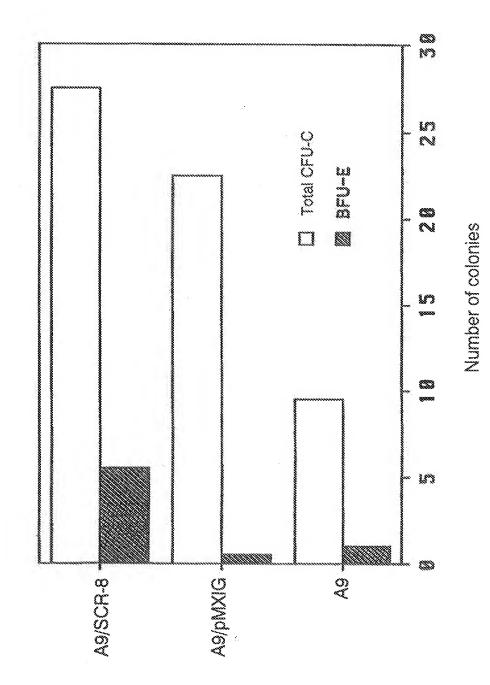


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Hig. 11